

Diversity and Species Composition of Bark and Ambrosia Beetles Captured Using Ethanol Baited Traps on Different Hosts in East Java, Indonesia

Hagus Tarno^{1,*}, Yogo Setiawan¹, Cindy B. Kusuma¹, Miftachul Fitriyah¹, Ahmad N. Hudan¹, Alvian P. Yawandika¹, Hanif A. Nasution¹, Ronauli Saragih¹, Achmad Praditya Yoga Bagasta¹, Zeng Wang², and Jianguo Wang³

¹*Environmental Entomology and nematology, Department Plant Pest and Disease, Faculty of Agriculture, Universitas Brawijaya. Jl. Veteran, Malang 65145, East Java, Indonesia.*

Correspondence: E-mail: h_gustarno@ub.ac.id (Tarno). Tel: +62-341-575843. Fax: 0341 560011.

E-mail: yogosetiawan07@gmail.com (Setiawan); cindybudikusuma5@gmail.com (Kusuma); ippetachul@gmail.com (Fitriyah); ahmadhudan@gmail.com (Hudan); dirgamasputraweka@gmail.com (Yawandika); hanifardiansyah87@gmail.com (Nasution); xxx@xxx (Saragih); yogabagasta@gmail.com (Yoga Bagasta)

²*Spice and Beverage Research Institute, Chinese Academy of Tropical Agricultural Sciences, Wanning, Hainan 571533, China. E-mail: sallywz618@163.com (Zeng)*

³*Laboratory of Invasion Biology, School of Agricultural Sciences, Jiangxi Agricultural University, Nanchang, Jiangxi 330045, China. E-mail: jgwang@jxau.edu.cn (Wang)*

(Received 23 June 2020 / Accepted 6 June 2021 / Published xx August 2021)

Communicated by Jen-Pan Huang

Bark and ambrosia beetles are a diverse group that cause widespread mortality of deciduous and coniferous trees. The present study aimed to investigate the species composition and richness of bark and ambrosia beetles in six species of plant hosts in East Java, Indonesia. Bark and ambrosia beetles were sampled using bottle traps baited with ethanol. Studies were conducted at two sites of monoculture and polyculture systems for each host plant species. At each site, 20 ethanol-baited traps were deployed on a linear transect along the forest. Six host tree species examined were used namely *Tectona grandis* (Teak), *Syzygium aromaticum* (Clove), *Swietenia mahagoni* (Mahogany), *Pinus merkusii* (Sumatran Pine), *Paraserianthes falcataria* (Moluccan Albizia), and *Mangifera indica* (Mango). The data were analyzed using R software. A total of 4823 beetles were collected, representing 26 ambrosia beetle and 8 bark beetle species. The abundance of bark and ambrosia beetles was significantly higher at the sites of *T. grandis* ($F = 13.88$, $P < 0.01$). *Xylosandrus crassiusculus* showed a strong attraction to the ethanol lure and was the dominant beetle species

(50.65% of the total number of individuals). The Shannon-Wiener diversity index of all beetles captured in this study was the highest in the *S. mahogany* polyculture (2.28) and the lowest in the *T. grandis* polyculture (0.47). According to Bray-Curtis analysis, the *T. grandis* monoculture and *T. grandis* polyculture had a high similarity value of bark and ambrosia beetle species composition (91% similar). There were no significant differences between two cultural systems of host plants in the composition of bark and ambrosia beetle species (ANOSIM, $R = -0.1537$, $P = 0.961$).

Key words: Ambrosia beetles, Cultural system, Ethanol-baited, Species richness, NMDS.

Citation: Tarno H, Setiawan Y, Kusuma CB, Fitriyah M, Nazaruddin AH, Yawandika AP, Nasution HA, Saragih R, Yoga Bagasta AP, Wang Z, Wang J. 2021. Diversity and Species Composition of Bark and Ambrosia Beetles Captured Using Ethanol Baited Traps on Different Hosts in East Java, Indonesia. Zool Stud 60:55.

BACKGROUND

Bark and ambrosia beetles are a diverse group that cause widespread mortality of deciduous and coniferous trees in forested and urban areas (Kühnholz et al. 2001; Oliver and Mannion 2001). The ambrosia beetle guild shows the lowest host specificity among whole herbivore guilds, and the bark beetle guild shows relatively high host specificity (Novotny et al. 2010). In general, ambrosia beetle females bore into the xylem and feed on symbiotic fungi, whereas bark beetles feed on the phloem of their host trees (Rabaglia et al. 2006).

Herbivorous insects have a degree of host specificity, from monophagous to polyphagous, and defensive capability, such as resistance to physical host defenses and its chemical compounds (Ødegaard et al. 2005; Agrawal 2007). The host specificity of herbivorous insects is one of the key predictors of patterns of biodiversity and has been widely used in the calculation of local species richness (Hamilton et al. 2010; Novotny et al. 2012). A recent study showed, in a tropical rainforest, that the model using host specificity is the best one for estimating species richness in herbivorous and nonherbivorous insect taxa (Basset et al. 2012).

In this study, six different plant species were used to estimate the species richness of bark and ambrosia beetles namely *Tectona grandis* (Teak), *Syzygium aromaticum* (Clove), *Swietenia mahagoni* (Mahogany), *Pinus merkusii* (Sumatran Pine), *Paraserianthes falcataria* (Moluccan Albizia), and *Mangifera indica* (Mango) in East Java, Indonesia. The present study also used bottle traps baited with ethanol. Traps with chemical attractants are commonly an effective control used for studying population dynamics, estimating species richness, predicting outbreaks, and mass

trapping to control pests (Burbano et al. 2012). Some studies have shown that a bottle trap baited with ethanol can be used to collect various species of bark and ambrosia beetle (Reding et al. 2011 2014; Galko et al. 2014). The collected information is needed to optimize detection, monitoring, and management programs for pest species in different plant hosts. The research objectives were to investigate the species composition and richness of bark and ambrosia beetles in six different plant hosts in East Java, Indonesia.

MATERIALS AND METHODS

Sampling Protocol

Bark and ambrosia beetles were sampled using bottle traps baited with ethanol (Fig. 1). Six different plant species were used in this study *i.e.*, *Tectona grandis* (Teak), *Syzygium aromaticum* (Clove), *Swietenia mahagoni* (Mahogany), *Pinus merkusii* (Sumatran Pine), *Paraserianthes falcataria* (Moluccan Albizia), and *Mangifera indica* (Mango). Diameter at breast height of selected sample trees ranged from 20 to 40 cm, except teak (> 40 cm). For each species, the study was divided into two sites based on cultural systems *i.e.* monoculture and polyculture. At each site, 20 ethanol-baited traps were deployed on a linear transect along forest edges and were separated by 20 m to reduce inter-trap interactions. The trap was made using a transparent bottle (volume = 1.5 L) with one window cut on the side and specimen container (containing soap solution) in the bellow part. It was baited with 95% ethanol. Each trap was ca. 7 cm in diameter (21.5 cm × 15 cm in size), attached on each trunk of sampled tree at approximately ca. 1.5 m above the ground (Fig. 1). Bark and ambrosia beetles were collected eight times at 3-day intervals. This research was conducted in Malang, Blitar, Mojokerto, Batu, Kediri, and Pasuruan from March to April 2017, and 2018, and from December 2018 to January 2019 (Table 1 and Fig. 2). Based on figure 2, the selected location of each site was depended on the suitability of cultural system of each plant host species.

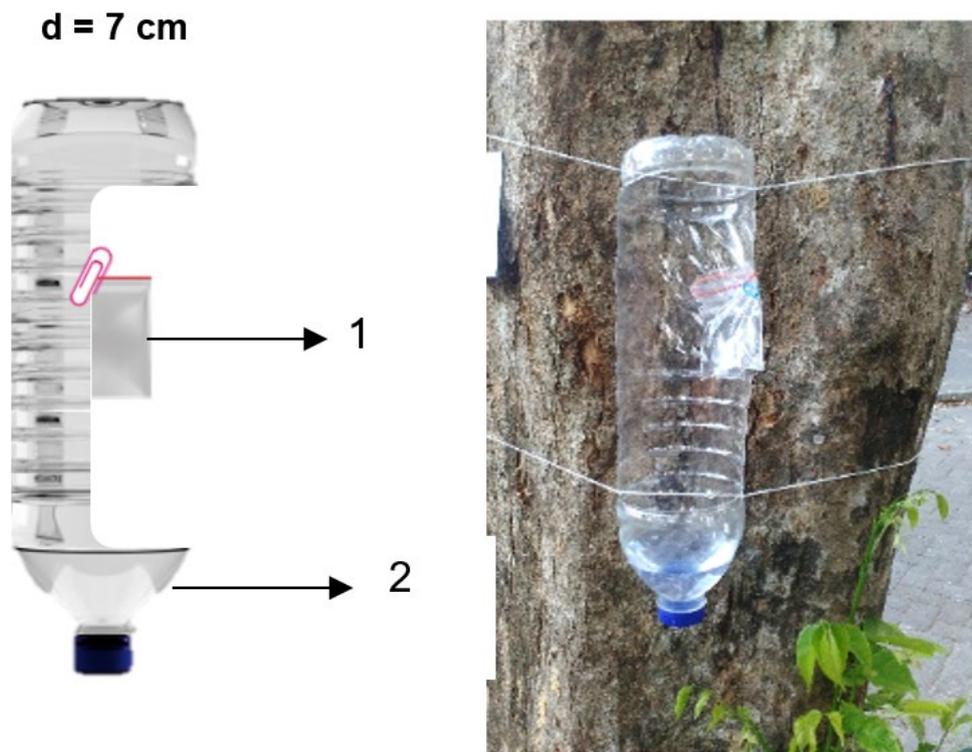


Fig. 1. Bottle trap baited with ethanol used in this study. 1. Plastic zipper bag containing 95% ethanol bait, and 2. Soap solution.

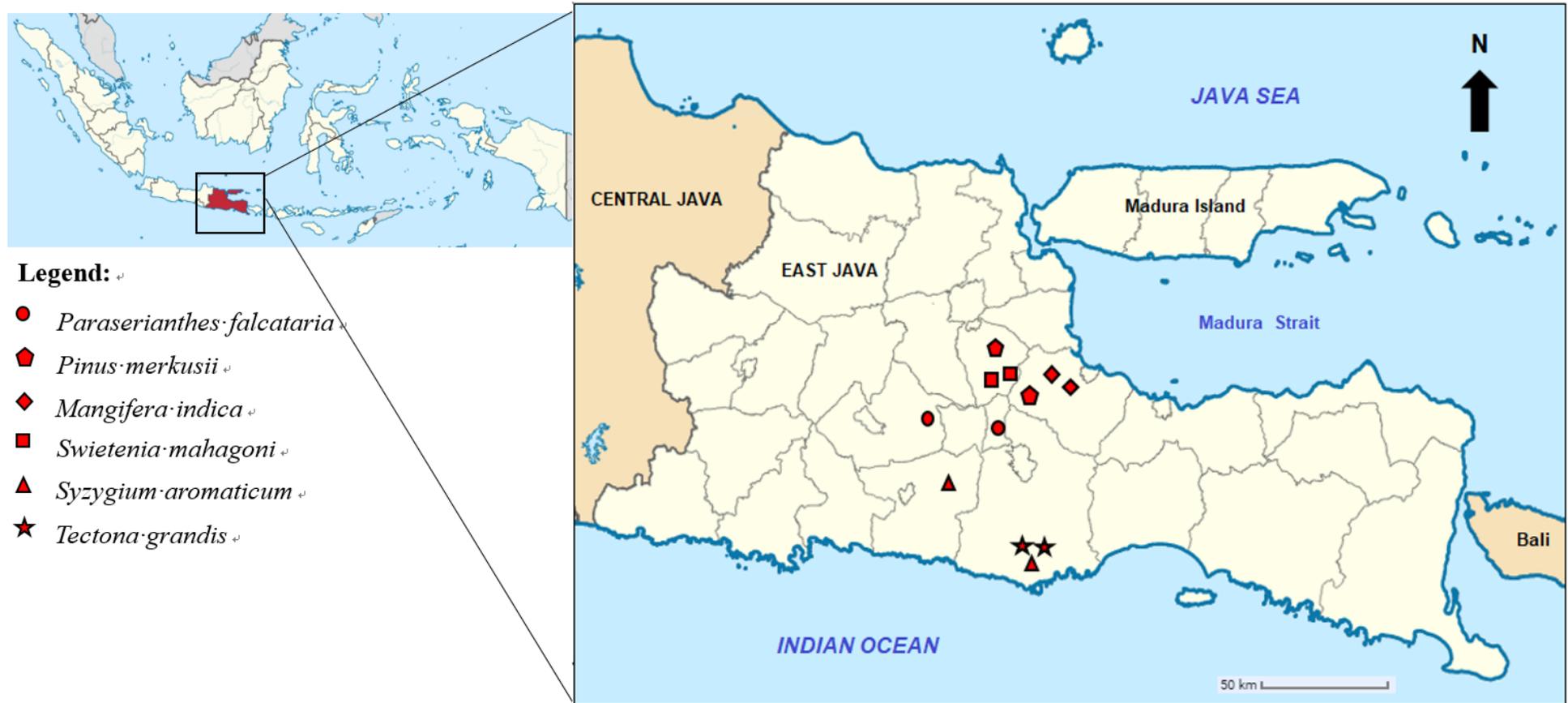


Fig. 2. Distribution of collection sites of each host plant species for bark and ambrosia beetles in East Java, Indonesia.

Preservation and Identification of Beetles

Bark and ambrosia beetles were preserved in 95% ethanol in small tubes. The bark and ambrosia beetle specimens were placed in the specimen bottle and labeled (date and site of observation). The identification of bark and ambrosia beetles was performed on the basis of morphological characters using an Olympus SZ51 stereo microscope. The identification of bark and ambrosia beetles was conducted at the Plant Pest Laboratory, Department of Plant Pests and Diseases, Faculty of Agriculture, Universitas Brawijaya. Bark and ambrosia beetles were identified using the Ambrosia beetle identification keys (Rabaglia et al. 2006; Wood 2007; Hulcr and Smith 2010).

Statistical Analysis

The populations of bark and ambrosia beetles in each host plant species were analyzed by using analysis of variance (ANOVA) ($P < 0.05$). Following significant results from ANOVA, the means were separated by Duncan's Multiple Range Test (DMRT) ($\alpha = 0.05\%$) using the R program version 3.3.3 with the vegan package *Agricolae*. The data were analyzed by using the Shannon-Wiener diversity index (H'), Evenness index (E), and Simpson Dominance Index ($1-D$) (Krebs 1999; Tarno et al. 2016). Bark and ambrosia beetle composition was compared between different host species based on the Bray-Curtis dissimilarity index and further analyzed using nonmetric multidimensional scaling (NMDS). All the data were analyzed by using the R program version 3.3.3 with the vegan package (Oksanen 2015; R Core Development Team 2019).

RESULTS

During the sampling period, a total of 4823 beetles, representing 26 species of ambrosia beetle and 8 species of bark beetle, were collected (Table 2). Among the six host trees examined, the abundance of bark and ambrosia beetles was significantly higher in *T. grandis* ($F = 13.88$, $P < 0.01$) (Fig. 3). In all of six different host plants investigated, the number of Ambrosia beetles exceeded the number of bark beetles. The former accounted for 95% (4586 individuals) of the total number of collected individuals, and the latter accounted for only 5% (237 individuals) of the total number of individuals. The ambrosia beetle *X. crassiuscullus* was the dominant species, with a total of 2443 individuals (50.65% of the total number of individuals collected) (Table 2).

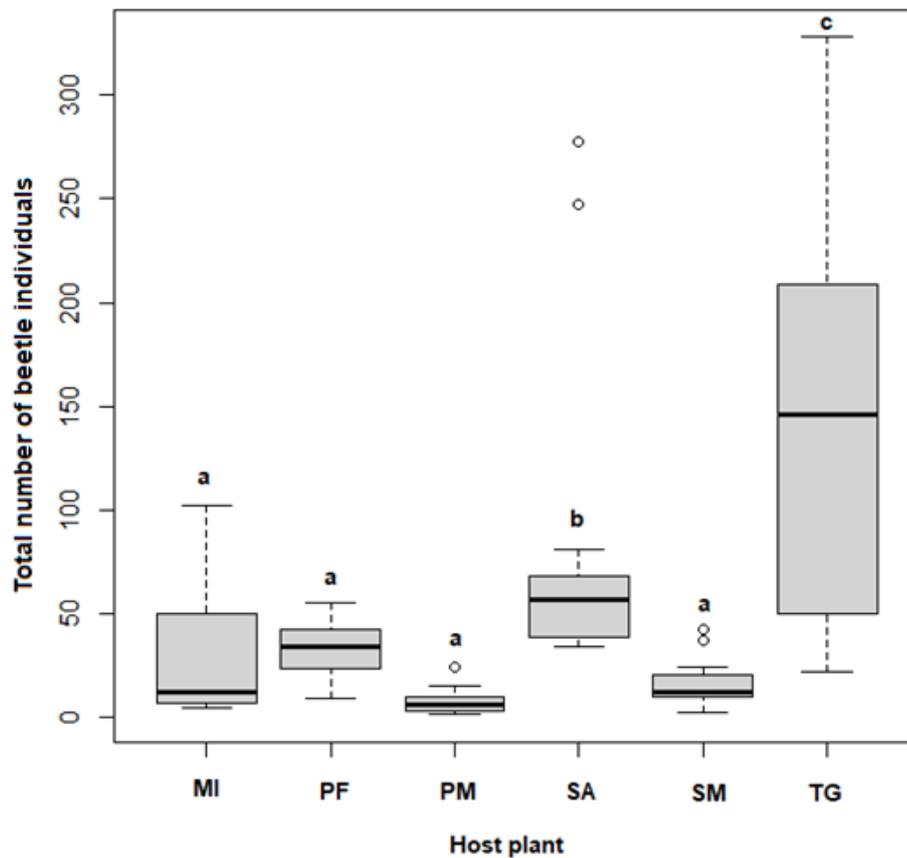


Fig. 3. The total number of bark and ambrosia beetles collected in six host plant species, including *Mangifera indica* (MI), *Paraserianthes falcataria* (PF), *Pinus merkusii* (PM), *Syzygium aromaticum* (SA), *Swietenia mahagoni* (SM), and *Tectona grandis* (TG).

The Shannon-Wiener diversity index of all beetles captured in this study was the greatest in the polyculture site of *S. mahogany* (2.28) and the lowest in the polyculture site of *T. grandis* (0.45) (Table 3). The species evenness index value for the polyculture site of *P. falcataria* (0.89) was higher than that for other hosts, and the lowest value was for the polyculture site of *T. grandis* (0.23) (Table 3). The Simpson's dominant index value was the highest (0.83) in the polyculture site of *T. grandis*, and the lowest (0.13) in the polyculture site of *S. mahogany* (Table 3).

NMDS ordination analysis showed the composition of bark and ambrosia beetle species in different host plants and culture systems (Fig. 3). NMDS ordination analysis showed no significant differences in the composition of bark and ambrosia beetle species between monoculture and polyculture (ANOSIM, $R = -0.1537$, $P = 0.961$) (Fig. 4). Based on Bray-Curtis analysis, the species composition of bark and ambrosia beetles on *T. grandis* trees were 91% similar between the site of polyculture and the site of monoculture systems (Fig. 5).

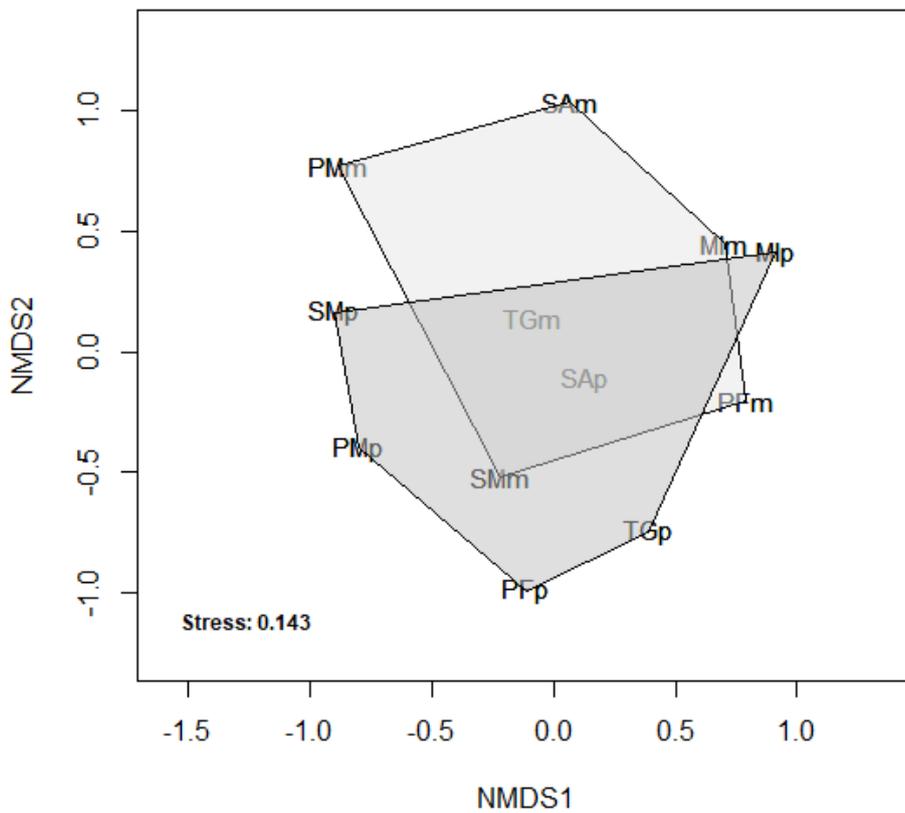


Fig. 4. Variation in bark and ambrosia beetle composition between study sites, in non-metric multidimensional scaling (NMDS) ordination (based on abundance data and a Bray-Curtis distance metric). The first & second letters indicate the host plant (TG: *Tectona grandis*, SA: *Syzygium aromaticum*, SM: *Swietenia mahagoni*. PM: *Pinus merkusii*, PF: *Paraserianthes falcataria*, MI: *Mangifera indica*), and the third letter indicates the cultural types (m: Monoculture, p: Polyculture).

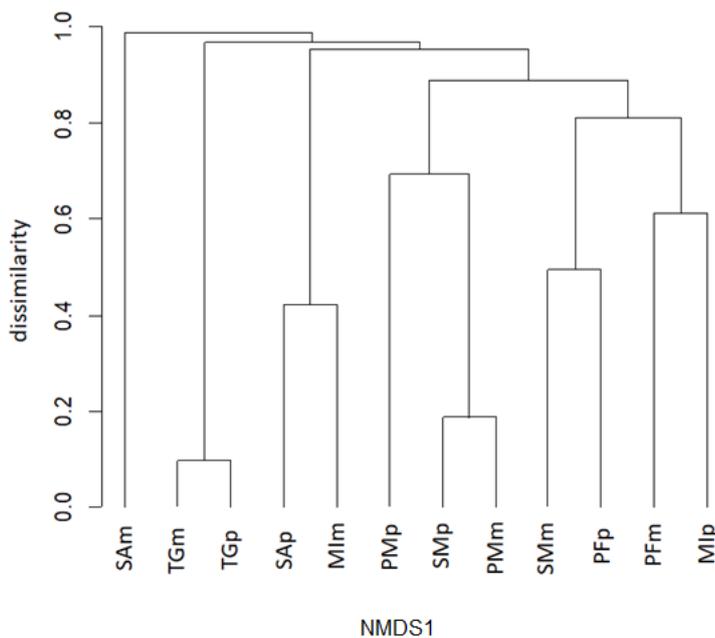


Fig. 5. Dissimilarity cluster between the host and the culture system in bark and ambrosia beetle species composition. The first & second letters indicate the host plant (TG: *Tectona grandis*, SA: *Syzygium aromaticum*, SM: *Swietenia mahagoni*. PM: *Pinus merkusii*, PF: *Paraserianthes falcataria*, MI: *Mangifera indica*), and the third letter indicates the cultural types (m: Monoculture, p: Polyculture).

DISCUSSION

Ambrosia and bark beetles in different plant host species in East Java were dominated by ambrosia beetle species, which constituted 95% of the total number of individuals and 5% of the total number of bark beetles in this study. Ambrosia beetles (Platypodidae and Scolytinae) also dominated in the teak forest and *Pterocarpus indicus* plant in the Malang District and Batu City, East Java (Tarno et al. 2014 2015; Setiawan et al. 2018). Five species of Platypodidae namely *Crossotarsus* sp., *Dinoplatypus* sp., *Dinoplatypus pallidus*, *Platypus solidus*, and *E. parallelus* were identified in this research. One of the species, *E. parallelus*, had previously been reported in Malang and Batu City, East Java (Tarno et al. 2014). *Xylosandrus crassiusculus* were the dominant species in this study, accounting for 50.65% of the total catch. Setiawan et al. (2018) reported that *X. crassiusculus* was the dominant species in polyculture and monoculture teak plant systems in the Malang District. Pennacchio et al. (2003) also reported that *X. crassiusculus* is a polyphagous species with various genera of plant hosts including forest trees, shrubs, and vines. It is also a pest in a variety of hosts, including 124 host and 48 families that occur mostly in tropical regions, including pine, cocoa, coffee, mahogany, rubber, tea, and teak (Horn and Horn 2006). Reding et al. (2011) also reported that bottle traps baited with ethanol lure captured *X. crassiusculus* effectively.

The present study showed that *T. grandis* had the highest total number of individuals. The species diversity of bark and ambrosia beetles was the greatest in *S. mahogany* baited with ethanol. According to Tarno et al. (2016), an index value between 1 and 3 is categorized as intermediate diversity, and the distribution of each species are also moderate. The species evenness index value for *P. falcataria* indicated the highest species evenness. According to Tarno et al. (2016), an index value between 0.75 and 1 is categorized as high species evenness and a stable species community. The Simpson's dominant index value of *P. falcataria* indicated low species dominance. According to Krebs (1999), when the index value is > 0.5 , there are no dominant species in the community. Species diversity can be influenced by several factors, including host specificity and mechanisms of plant resistance to herbivorous insects. In this study, the diversity of ambrosia beetles in various host species differed depending on the resistance mechanism of each plant species. The host specificity of herbivorous insects is one of the key predictors of patterns of biodiversity (Novotny et al. 2002). Host plants had variation in defensive capability, *i.e.*, chemical compounds and physical defenses, such as lignin or wood toughness (Agrawal 2007). The lowest value of diversity was *T. grandis*, because this plant had natural resistance properties to insects. Ngee et al. (2004) reported that teak was a less preferred species against the native pests *Coptotermes*, *Microceratotermes*, *Globitermes*, and *Macrotermes* termites in a preference test. The chemical components provide a

good indication of the natural durability of teak, and the composition of extractives of teak wood was reported to be quite complex (Yamamoto et al. 1998). According to Lukmandaru and Takahashi (2008), tectoquinone (2-methylanthraquinone), and the n-hexane-extractible content has insecticidal properties, conferring insect resistance.

In this study, the *T. grandis* monoculture and *T. grandis* polyculture had a high similarity value. Differences in species composition between plant host species and *X. affinis* were found in all sites. In this study, each host plant species had different locations, which differed in available resources, and altitude. At the study sites, *S. aromaticum* and *M. indica* had the lowest altitude. Warmer temperatures and climate changes at lower elevations may lengthen the flight activity period and increase the number of generations produced per year, thus inducing beetles to migrate to higher elevations (Bale et al. 2002). *Xyleborus affinis* was found in all sites, thus it had the broadest host range. *Xyleborus affinis* is extremely polyphagous and has a known host range of 248 species, angiosperms as well as gymnosperms (Wood 1982). Although it is among the most widespread and common ambrosia beetles in forested areas around the world (Sobel et al. 2015). Steininger et al. (2015) also reported that *X. affinis* was only weakly attracted to ethanol, the most commonly used lure for ambrosia beetle monitoring.

Based on NMDS ordination analysis, no significant differences in the composition of bark and ambrosia beetle species between monoculture and polyculture describes that both cultural systems are similar. Reed and Muzika (2010) reported that different forms of forest management may not modify the ambrosia beetle community. Setiawan et al. (2018) also mentioned that in teak forest, monoculture and polyculture systems had the same diversity categories based on ambrosia beetle abundant. Previous studies provide evidence about similarity on two cultural systems of six different host plants related to the ambrosia and bark beetles abundant in East Java, Indonesia.

CONCLUSIONS

A total of 4823 beetles were collected, representing 26 ambrosia beetle and 8 bark beetle species. The abundance of bark and ambrosia beetles was significantly higher in *T. grandis*. *X. crassiuscullus* showed a strong attraction to the ethanol lure and was the dominant species. The Shannon-Wiener diversity index of all beetles captured in this study was the greatest in the *S. mahogany* polyculture and the lowest in the *T. grandis* polyculture. According to Bray-Curtis analysis, the *T. grandis* monoculture and *T. grandis* polyculture had a higher similarity value of bark and ambrosia beetle species composition than cultures collected at other sites. There were no significant differences between culture systems in the composition of bark and ambrosia beetle species.

Acknowledgment: This research was supported partly by the China Association for Science and Technology “China-Indonesia bilateral scientific and technological exchanges on disease and insect pest control technology of tropical fragrant beverage crops”(2020). In addition, authors thank You Li and Andrew Johnson from the University of Florida, USA for their help to confirm the identification of the beetles.

Authors’ contribution: Hagus Tarno designed the study, analyzed the specimen and finalized the manuscript. Cindy Budi Kusuma, Miftachul Fitriyah, Ahmad Hudan Nazaruddin, Alvian Putra Yawandika, Hanif Ardiansyah Nasution, Ronauli Saragih, and Achmad Praditya Yoga Bagasta performed the field work. Yogo Setiawan performed statistical analyses and wrote the manuscript draft. Zeng Wang, and Jianguo Wang performed the statistical analyses.

Competing interest: The authors declare that they have no conflict of interests.

Availability of data and materials: Data available.

Consent for publication: Not applicable.

Ethics approval consent to participate: Not applicable.

REFERENCES

- Agrawal AA. 2007. Macroevolution of plant defense strategies. *Trends Ecol Evol* **22(2)**:103–109. doi:10.1016/j.tree.2006.10.012.
- Bale JS, Masters GJ, Hodkinson ID, Awmack C, Bezemer TM. 2002. Herbivory in global climate change research: direct effects of rising temperature on insect herbivores. *Glob Change Biol* **8(1)**:1–16. doi:10.1046/j.1365-2486.2002.00451.x.
- Basset Y, Cizek L, Cuenoud P, Didham RK, Guilhaumon F. 2012. Arthropod diversity in a tropical forest. *Science* **338(6113)**:1481–1484. doi:10.1126/science.1226727.
- Burbano EG, Wright MG, Gillette NE, Mori A, Dudley N. 2012. Efficacy of Traps, Lures, and Repellents for *Xylosandrus compactus* (Coleoptera: Curculionidae) and other Ambrosia Beetles on Coffea arabica Plantations and Acacia koa Nurseries in Hawaii. *Environ Entomol* **41(1)**:133–140. doi:10.1603/EN11112.
- Galko J, Nikolov C, Kimoto T, Kunca A, Gubka A. 2014. Attraction of ambrosia beetles to ethanol baited traps in a Slovakian oak forest. *Biologia (Bratisl)* **69(10)**:1376–1383.

- Hamilton AJ, Basset Y, Benke KK, Grimbacher PS, Miller SE. 2010. Quantifying uncertainty in estimation of tropical arthropod species richness. *Am Nat* **176(1)**:90–95. doi:10.1086/652998.
- Horn S, Horn GN. 2006. New host record for the Asian ambrosia beetle, *Xylosandrus crassiusculus* (Wlutschulsky) (Coleoptera: Curculionidae). *J Entomol Sci* **41(1)**:90–91.
- Hulcr J, Smith S. 2010. Xyleborini ambrosia beetles: an identification tool to the world genera.
- Krebs CJ. 1999. *Ecological Methodology*. 2nd Edition, Benjamin Cummings, Menlo Park, 620 p.
- Kühnholz S, Borden JH, Uzunovic A. 2001. Secondary ambrosia beetles in apparently healthy trees: adaptations, potential causes and suggested research. *Integr Pest Manag Rev* **6(3–4)**:209–219. doi:10.1023/A:1025702930580.
- Lukmandaru G, Takahashi K. 2008. Variation in the natural termite resistance of teak (*Tectona grandis* Linn. fil.) wood as a function of tree age. *Ann For Sci* **65(7)**:708. doi:10.1051/forest:2008047.
- Ngee PS, Tashiro A, Yoshimura T, Jaal Z, Lee CY. 2004. Wood preference of selected Malaysian subterranean termites (Isoptera: Rhinotermitidae, Termitidae). *Sociobiology* **43(3)**:535–550. doi:10.1007/BF00988182.
- Novotny V, Basset Y, Miller SE, Weiblen GD, Bremer B et al. 2002. Low host specificity of herbivorous insects in a tropical forest. *Nature* **416(6883)**:841–844. doi:10.1038/416841a.
- Novotny V, Miller SE, Baje L, Balagawi S, Basset Y. 2010. Guild-specific patterns of species richness and host specialization in plant-herbivore food webs from a tropical forest. *J Anim Ecol* **79(6)**:1193–1203. doi:10.1111/j.1365-2656.2010.01728.x.
- Novotny V, Miller SE, Hrcek J, Baje L, Basset Y. 2012. Insects on plants: explaining the paradox of low diversity within specialist herbivore guilds. *Am Nat* **179(3)**:351–362. doi:10.1086/664082.
- Ødegaard F, Diserud OH, Østbye K. 2005. The importance of plant relatedness for host utilization among phytophagous insects. *Ecol Lett* **8(6)**:612–617. doi:10.1111/j.1461-0248.2005.00758.x.
- Oksanen J. 2015. Vegan: ecological diversity. **1(2)**:1–12. doi:10.1029/2006JF000545 (not this article. wrong year. should ask the authors).
- Oliver JB, Mannion CM. 2001. Ambrosia beetle (Coleoptera: Scolytidae) species attacking chestnut and captured in ethanol-baited traps in middle Tennessee. *Environ Entomol* **30(5)**:909–918. doi:10.1603/0046-225X-30.5.909.
- Pennacchio F, Roversi PF, Francardi V, Gatti E. 2003. *Xylosandrus crassiusculus* (Motschulsky) A Bark Beetle New to Europe (Coleoptera Scolytidae). *Redia* **86(2)**:77–80.
- R Core Development Team. 2019. R: a language and environment for statistical computing.
- Rabaglia RJ, Dole SA, Cognato AI. 2006. Review of American Xyleborina (Coleoptera: Curculionidae: Scolytinae) occurring north of Mexico, with an illustrated key. *Ann Entomol*

- Reding ME, Schultz PB, Ranger CM, Oliver JB. 2011. Optimizing ethanol-baited traps for monitoring damaging ambrosia beetles (Coleoptera: Curculionidae, Scolytinae) in ornamental nurseries. *J Econ Entomol* **104(6)**:2017–2024. doi:10.1603/EC11119.
- Reed SE, Muzika RM. 2010. The influence of forest stand and site characteristics on the composition of exotic dominated ambrosia beetle communities (Coleoptera: Curculionidae: Scolytinae). *Environ Entomol* **39(5)**:1482–1491. doi:10.1603/EN09374.
- Setiawan Y, Rachmawati R, Tarno H. 2018. Diversity of ambrosia beetles (Coleoptera: Scolytidae) on teak forest in Malang District, East Java, Indonesia. *Biodiversitas J Biol Divers* **19(5)**:1783–1790. doi:10.13057/biodiv/d190528.
- Sobel L, Lucky A, Hulcr J. 2015. An Ambrosia Beetle *Xyleborus affinis* Eichhoff, 1868 (Insecta: Coleoptera: Curculionidae: Scolytinae). *Entomology Nematol Dep. UF*: pp. 1–5.
- Steininger MS, Hulcr J, Šigut M, Lucky A. 2015. Simple and efficient trap for bark and ambrosia beetles (Coleoptera: Curculionidae) to facilitate invasive species monitoring and citizen involvement. *J Econ Entomol* **108(3)**:1115–1123. doi:10.1093/jee/tov014.
- Tarno H, Septia ED, Aini LQ. 2016. Microbial community associated with ambrosia beetle, *Euplatypus parallelus* on sonokembang, *Pterocarpus indicus* in Malang. *Agrivita, J Agric Sci* **38(3)**:312–320.
- Tarno H, Suprpto H, Himawan T. 2014. First record of ambrosia beetle (*Euplatypus paralellus* fabricius) infestation on sonokembang (*Pterocarpus indicus* willd.) from Malang Indonesia. *Agrivita, J Agric Sci* **36(2)**:189–200. doi:10.17503/Agrivita-2014-36-2-p189-200.
- Tarno H, Suprpto H, Himawan T. 2015. New record of the ambrosia beetle, *Treptoplatypus micrurus* schedl. Attack on sonokembang (*Pterocarpus indicus* Willd.) in Batu, Indonesia. *Agrivita* **37(3)**:220–225. doi:10.17503/Agrivita-2015-37-3-p220-225.
- Wood SL. 1982. The bark and ambrosia beetles of North and Central America (Coleoptera: Scolytidae), a taxonomic monograph. *Gt Basin Nat Mem* **6**:1–1359. Available at <https://www.biodiversitylibrary.org/part/248626>. Accessed dd mm year.
- Wood SL. 2007. *Bark and Ambrosia Beetles*. Print and Mail Production Center, Provo, Utah USA.
- Yamamoto K, Simatupang MH, Hashim R. 1998. Caoutchouc in teak wood (*Tectona grandis* L.f.): formation, location, influence on sunlight irradiation, hydrophobicity and decay resistance. *Holz als Roh - und Werkst* **56(3)**:201–209. doi:10.1007/s001070050299.