

## Dry Season Feeding Profiles of a Characiformes Assemblage in a Brazilian Tropical Stream

Elioenai da Silva Oliveira<sup>1,2,\*</sup>, Josie South<sup>3</sup>, Lucas de Oliveira Vieira<sup>1,2</sup>, and Felipe Polivanov Ottoni<sup>1,2,4</sup>

<sup>1</sup>Programa de Pós-graduação em Biodiversidade e Biotecnologia da Amazônia Legal, Universidade Federal do Maranhão, Av. dos Portugueses, 1966, Cidade Universitária Dom Delgado, 65080-805, São Luís, MA, Brazil.

\*Correspondence: E-mail: oliveiraelioenai@hotmail.com (da Silva Oliveira)

E-mail: lucasolivier070@gmail.com (de Oliveira Vieira); fpottoni@gmail.com (Ottoni)

<sup>2</sup>Laboratório de Sistemática e Ecologia de Organismos Aquáticos, Universidade Federal do Maranhão, BR-222, KM 04, Boa Vista, 65500-000, Chapadinha, Maranhão, Brazil

<sup>3</sup>Water@Leeds, School of Biology, Faculty of Biological Sciences, University of Leeds, Leeds LS2 9JT, United Kingdom. E-mail: J.South@leeds.ac.uk (South)

<sup>4</sup>Programa de Pós-graduação em Ciências Ambientais, Universidade Federal do Maranhão, BR-222, KM 04, Boa Vista, 65500-000, Chapadinha, Maranhão, Brazil

<sup>5</sup>NRF-South African Institute for Aquatic Biodiversity (NRF-SAIAB), P. Bag 1015, Makhanda 6140, South Africa

(Received 5 June 2025 / Accepted 22 December 2025 / Published -- 2026)

Communicated by Jen-Pen Huang

Trophic interactions between fish and their resources depends on resource availability and interspecific competition. To understand dry season trophic profiles of a speciose Characiformes assemblage we performed stomach content analysis to describe diet and determine levels of niche partitioning and morphological adaptations among eight Characiformes species in the dry season in Mata de Itamacaoca, Chapadinha Municipality, State of Maranhão, northeastern Brazil. Insectivory dominated most diets, with *Astyanax* cf. *bimaculatus* and *Characidium* cf. *bimaculatum* exhibiting the broadest niches. Specialization occurred in *Curimatopsis* cf. *cryptica* (85.07% plant material) and there was significant dietary segregation with indicator species analysis linking *Astyanax* cf. *bimaculatus* to piscivory and *Knodus guajajara* to vermicivory. Pianka index showed extreme niche overlap variations, with the highest overlap between *Bario oligolepis* and *Characidium* cf. *bimaculatum* (1.68), and between *Astyanax* cf. *bimaculatus* and *Nannostomus beckfordi* (1.64). Morphological PCA associated traits with feeding strategies: caudal fin length (*Astyanax* cf. *bimaculatus*), body depth (*Curimatopsis* cf. *cryptica*), and oral gape width (*Bario oligolepis*). Mixed models confirmed insects and plant material with a marginally significant effect as key drivers of dietary variation. Therefore, the assemblage shows high niche overlap combined with diverse trophic profiles. Results presented here demonstrate how dry season resource scarcity promotes trophic divergence via morphological specialization, with generalists (*Astyanax* cf.

*bimaculatus*) coexisting with specialists through niche partitioning, which is critical for conservation in this threatened urban-protected area.

**Keywords:** Morphological adaptations, Neotropical fishes, Resource partitioning, Seasonality, Trophic ecology

Citation: da Silva Oliveira E, South J, de Oliveira Vieira L, Ottoni FP. 2026. Dry season feeding profiles of a Characiformes assemblage in a Brazilian tropical stream. Zool Stud 65:02.

## BACKGROUND

Neotropical aquatic ecosystems harbor one of the most diverse ichthyofaunas on the planet (Albert et al. 2020; Tonella et al. 2022), where Characiformes play a fundamental role in structuring trophic networks (Barreto and Aranha 2006; Silva-Camacho et al. 2014; Meira et al. 2022; Oliveira et al. 2024). In seasonal environments, hydrological variation acts as an environmental filter, shaping patterns of trophic and morphological adaptations (Junk et al. 1989; Correa and Winemiller 2014; Duarte et al. 2022). Previous studies have shown that the dry season imposes critical constraints on resource availability, leading to increased interspecific competition (Prejs and Prejs 1987), the emergence of distinct morphological strategies (Gomiero et al. 2010), and dietary specialization (Novakowski et al. 2008). Although trophic segregation has been highlighted as the primary mechanism structuring fish assemblages (Ross 1986), this dynamic may vary according to local conditions, including dry season factors (Bouton et al. 1997). However, gaps remain in understanding the mechanisms that allow the coexistence of multiple sympatric species under such extreme conditions (Ross 1986; Neves et al. 2018).

Aquatic environments are generally strongly influenced by seasonal periods and flood pulse dynamics (Junk et al. 1989; Pazin et al. 2006; Espírito-Santo and Zuanon 2017). As flood peaks reach their maximum and the system transitions into the dry season, periods that are becoming increasingly pronounced, there is a progressive decline in turbidity, resource availability, flow velocity, and water level (Alho and Silva 2012). These abiotic changes result in significant transformations in fish assemblages (Saint-Paul et al. 2000). While some species exhibit expansion and contraction dynamics aligned with dry season reproduction, others persist throughout the entire hydrological cycle (Fialho et al. 2008; Arthington and Balcombe 2011; Fitzgerald et al. 2018). Dry season variation, particularly in tropical regions, plays a crucial role in shaping food resource availability and structuring trophic networks (Medeiros et al. 2014; Pelage et al. 2022; Londe et al.

2024). During the dry season, reduced water volume can lead to increased population density and the concentration of organisms in remnant habitats, intensifying ecological interactions such as competition and predation (Duarte et al. 2022). This scenario can directly impact niche partitioning, leading to shifts in dietary composition and potential trophic displacements among sympatric species (Silva-Camacho et al. 2014; Bloomfield et al. 2022; De Andrade et al. 2024).

In the context of dry season persistence, intraspecific morphological variation becomes a crucial factor for fish survival in stochastic ecosystems, as species evolve in response to persistent hydrological regimes (Poff and Ward 1989; Lytle and Poff 2004). Morphological adaptations and diversity can confer specializations to specific environmental parameters, thereby increasing survival among cohorts (Langerhans and Reznick 2010). Morphological theory predicts that coexistence in restrictive environments is mediated by three main mechanisms: (a) divergence in functional traits (Winemiller 1991), (b) behavioral plasticity (Correa and Winemiller 2014), and (c) temporal resource partitioning (Ross 1986). However, the application of these principles to small Characiformes assemblages in seasonal microhabitats remains insufficiently tested. Studies in analogous systems suggest that body and oral apparatus morphology explain up to 80% of the variation in resource use (Neves et al. 2018; Duarte et al. 2022), but these patterns may differ significantly in fragmented environments such as the Mata de Itamacaoca.

The order Characiformes is one of the most diverse among Neotropical fishes, comprising approximately 1,700 described species (Reis et al. 2016) and encompassing a wide range of feeding habits, from herbivores and detritivores to carnivores and piscivores (Barbosa et al. 2017; Burns and Sidlauskas 2019). This functional diversity grants these fishes a crucial role in mediating energy and matter flow in aquatic ecosystems, directly influencing the availability and renewal of trophic resources (Burns and Sidlauskas 2019; Burns 2021; Burns et al. 2024). Moreover, their abundance and distribution across different habitats make them ideal models for investigating trophic interactions and adaptive strategies in dry season environments (Burns and Sidlauskas 2019; Burns et al. 2024). Trophic ecology among Characiformes species is often associated with morphological differences, particularly in mouth shape, dentition, and digestive tract structure (Silva-Camacho et al. 2014; Benone et al. 2020; Burns 2021; Meira et al. 2022). Specialized morphological traits enable differential exploitation of available resources (Sibbing and Nagelkerke 2000; Bower and Winemiller 2019), reducing dietary overlap (Mise et al. 2013) and promoting the coexistence of multiple species within the same environment (Oliveira et al. 2024; Oliveira et al., 2025). In environments influenced by seasonal hydrological regimes, these adaptations can be essential for species survival, allowing diversification of feeding strategies as resource availability fluctuates throughout the hydrological cycle (Porter et al. 2022; Bloomfield et al. 2022; De Andrade et al. 2024).

The Munim River Basin (16,000 km<sup>2</sup>), an important hydrographic system of Maranhão (Koerber et al. 2022), which is located in a transitional zone between the Amazon and Cerrado biomes (NuGeo 2016), harboring a still understudied ichthyofauna (Abreu et al. 2019; Vieira et al. 2023). Within this context, the Mata de Itamacaoca stands out as a unique ecological enclave embedded within an urban matrix (Oliveira et al. 2020), sustaining a diverse assemblage of small Characiformes (Oliveira et al. 2020), characterized by significant morphological and trophic overlap (Oliveira et al. 2024). The coexistence of functionally similar species in a seasonally dynamic environment suggests (i) the presence of sophisticated resource partitioning mechanisms (Burns and Sidlauskas 2019) and (ii) an increased vulnerability to anthropogenic disturbances (Daufresne and Boet 2007). Although preliminary studies have identified trophic segregation patterns (Oliveira et al. 2024), possible mechanisms are unexplored as these studies combined both wet and dry season than accounting for increased resource abundance in the wet season. Thus, dry-season ecological processes in the Munim River Basin remain poorly understood, particularly regarding how seasonal reductions in water volume and resource availability shape trophic interactions among fish species (Junk et al. 1989; Lytle and Poff 2004; Correa and Winemiller 2014).

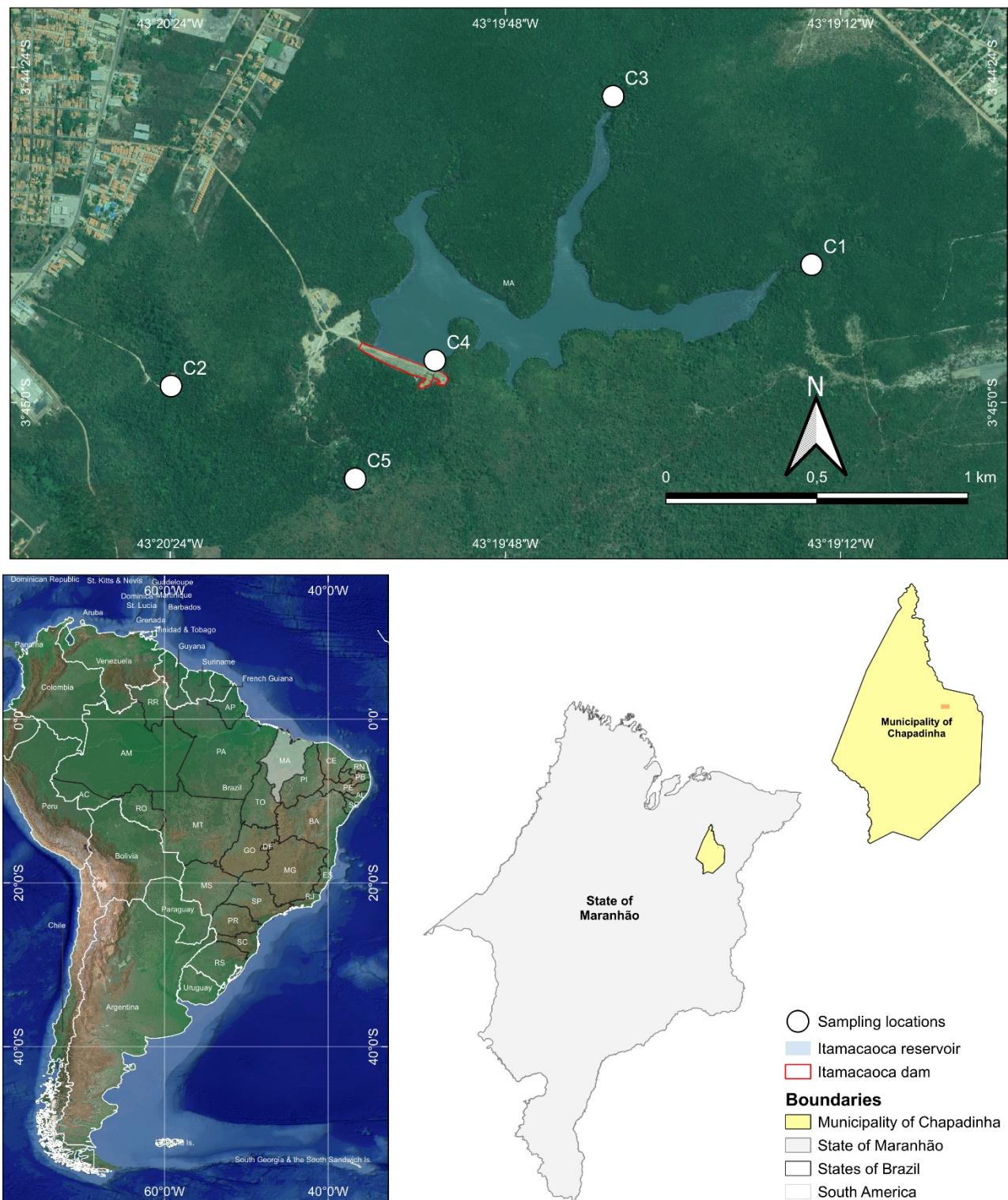
Given the above, this study aims to investigate the dietary composition and trophic structure of Characiformes species in the Mata de Itamacaoca during the dry season through stomach content analysis, correlating it with food resource availability and species' morphological adaptations. Specifically, we seek to: (1) describe dietary composition and identify the main food items consumed based on stomach content analysis, (2) assess patterns of overlap and segregation in resource use among species, (3) examine the relationship between morphological attributes and dietary preferences, and (4) discuss the ecological implications of resource partitioning and interspecific competition.

## MATERIALS AND METHODS

### Study area and sampling methodology

This study was conducted in the Mata de Itamacaoca (middle Munim River Basin), a protected urban fragment (460 ha) within the Cerrado biome 03°44'45.2"S 43°19'15.1"W; ~90 m elevation), located in the Chapadinha municipality, State of Maranhão, northeastern Brazil (Fig. 1, Table 1). Mata de Itamacaoca encompasses a diverse array of microhabitats, including riparian forests, gallery forests, and perennial streams that collectively support a rich biodiversity

representative of the Cerrado biome (Silva et al. 2008; Oliveira et al. 2020). The vegetation consists primarily of closed-canopy formations with trees exceeding 10 meters in height, particularly around springs and water bodies, which are essential for maintaining local water supplies (Silva et al. 2008). The area was officially designated as an Area of Relevant Ecological Interest (Decreto N° 05/2018) due to its critical role in watershed protection, microclimate regulation, and the conservation of regional biodiversity (Silva et al. 2008). Despite its protected status, the reserve faces increasing anthropogenic pressures, including illegal resource extraction (e.g., timber, fish, and game), agricultural burning practices, urban encroachment, and inadequate enforcement of conservation measures (Oliveira et al. 2020). These threats have significantly affected both the hydrological dynamics of the reservoir system and the conservation status of aquatic biodiversity in recent years. The area's high accessibility and complete urban encroachment make it particularly vulnerable to such disturbances, despite its recognized ecological importance for regional water supply and climate regulation (Oliveira et al. 2020).



**Fig. 1.** Location of the collecting sites (C1-C5) distributed across the Mata de Itamacaoca, Chapadinha municipality, State of Maranhão, northeastern Brazil.

**Table 1.** Description of the collecting sites, including coordinates and habitat characteristics, in Mata de Itamacaoca, Chapadinha, Maranhão, Brazil

Collecting Site	Coordinates	Habitat Characteristics
C1	3°44'45.20"S 43°19'15.10"W	Stream near a spring, surrounded by gallery and riparian forest, in Mata de Itamacaoca, Chapadinha, Maranhão. Sampling covered ~200 meters of the watercourse.
C2	3°44'58.24"S 43°20'23.91"W	Stream in the Repouso do Guerreiro area, within Mata de Itamacaoca, Chapadinha, Maranhão.
C3	3°44'27.1"S 43°19'36.4"W	Stream near a natural water source, with gallery and riparian forest, in Mata de Itamacaoca, Chapadinha, Maranhão.
C4	3°44'55.16"S 43°19'57.10"W	Itamacaoca Dam, located in Chapadinha, Maranhão.
C5	3°45'8.20"S 43°20'4.13"W	Stream downstream of the dam, within Mata de Itamacaoca, Chapadinha, Maranhão.

The regional climate exhibits strong seasonality, with a well-defined dry season lasting five to six months (July to November/December), characterized by significant water deficits (150–300 mm), followed by an equally distinct rainy season from January to May/June, with peak precipitation occurring between February and March (Passos et al. 2016; IMESC 2021). This marked seasonal variation may create dynamic environmental conditions that profoundly influence the aquatic ecosystems within the protected area.

Sampling was conducted during the dry season (from July to December 2019) at five previously established collecting sites (C1-C5) distributed across the Mata de Itamacaoca within the middle Munim River Basin (Fig. 1, Table 1). All sampling procedures were authorized under SISBIO permit N° 64415. Because the study involved only the collection of wild fish specimens for taxonomic and ecological analyses, it did not require approval from an Institutional Animal Care and Use Committee (CEUA). These sites included both natural stream sections and one dam-impacted area (C4), as described in Oliveira et al. (2020) (Fig. 1, Table 1). Fish collections were performed using standardized techniques with dip nets (80 cm × 54 cm, 2 mm mesh) and trail nets (240 cm × 100 cm, 2 mm mesh) following the methodology of Souza and Auricchio (2002). All collection procedures adhered to animal welfare guidelines (Underwood and Anthony 2020), with specimens euthanized in a solution of ethyl-3-amino-benzoate-methanesulfonate (MS-222; 250 mg/L) until cessation of opercular movement. Following euthanasia, specimens were initially preserved in 10% formalin and subsequently transferred to 70% ethanol after 10-15 days for long-term storage. Voucher specimens are housed at the Coleção Ictiológica do Centro de Ciências Agrárias e Ambientais (CICCAA) of the Universidade Federal do Maranhão (UFMA); the complete information spreadsheets are provided in table S1. This sampling design-maintained consistency with previous studies in the area while specifically targeting the dry season to investigate trophic and morphological adaptations under seasonal stress conditions.

## Fish Identification

Fish were identified to the lowest possible taxonomic level, based on specific literature for each taxonomic group. Species names, authorship and year of description, geographical distribution, taxonomic classification, as well as other additional information were checked in Fricke et al. (2025a, b).

## Stomach content analyses

Only adult individuals were included in all analyses to avoid ontogenetic effects on trophic composition and morphological traits (Winemiller 1991; Gerking 1994). This was confirmed by examining standard length (SL) ranges for each species (Table 2), which consistently corresponded to adult size classes reported in the literature. We analyzed the dietary composition of 173 specimens belonging to eight Characiformes species: *Astyanax* cf. *bimaculatus* ( $n = 26$ ; Acestrorhaphidae), *Characidium* cf. *bimaculatum* ( $n = 27$ ; Crenuchidae), *Curimatopsis* cf. *cryptica* ( $n = 23$ ; Curimatidae), *Holopristis* cf. *ocellifera* [*Hemigrammus* sp. 1 *sensu* Oliveira et al. (2020)] ( $n = 30$ ; Acestrorhaphidae), *Hyphessobrycon* *piorskii* Guimarães, Brito, Feitosa, Carvalho-Costa & Ottoni 2018 ( $n = 16$ ; Acestrorhaphidae), *Knodus* *guajajara* Aguiar, Brito, Ottoni & Guimarães 2022 [*Knodus* *victoriae* (Steindachner, 1907) *sensu* Oliveira et al. (2020)] ( $n = 10$ ; Stevardiidae), *Bario* *oligolepis* (Günther 1864) ( $n = 11$ ; Acestrorhaphidae), and *Nannostomus beckfordi* Günther, 1872 ( $n = 30$ ; Lebiasinidae) (Table S1, Table 2). An ideal sample size of 30 individuals per species was initially established to standardize comparisons. However, some species did not reach this number due to their low abundance in the sampled environment during the dry season. Despite this limitation, the available sample sizes were considered adequate for descriptive dietary and morphological analyses.

**Table 2.** Standard length (SL) variation of Characiformes fishes sampled in Mata de Itamacaoca during the dry season of 2019. Values represent: N = sample size per species, size range (min-max), mean  $\pm$  standard deviation (SD), and median SL in millimeters

Family	Species	N	SL Range (mm)	SL Mean $\pm$ SD (mm)	SL Median (mm)
Acestrorhaphidae	<i>Astyanax</i> cf. <i>bimaculatus</i>	26	27.5-76.96	53.35 $\pm$ 9.1	53.86
	<i>Bario</i> <i>oligolepis</i>	11	45.86-68.44	52.65 $\pm$ 6.02	51.73
	<i>Holopristis</i> cf. <i>ocellifera</i>	30	25.09-34.15	30.76 $\pm$ 2.08	31.17
	<i>Hyphessobrycon</i> <i>piorskii</i>	16	21.02-28.5	25.2 $\pm$ 2.01	25.26
Crenuchidae	<i>Characidium</i> cf. <i>Bimaculatum</i>	27	22.91-27.55	24.99 $\pm$ 1.07	24.77
Curimatidae	<i>Curimatopsis</i> cf. <i>cryptica</i>	23	30.48-40.42	33.57 $\pm$ 2.94	32.63
Lebiasinidae	<i>Nannostomus beckfordi</i>	30	25.83-29.8	27.75 $\pm$ 1.04	27.89
Stvardiidae	<i>Knodus</i> <i>guajajara</i>	10	23.88-36.48	30.00 $\pm$ 4.57	30.56

To achieve this, we removed the stomach and intestine of each individual and placed the digestive contents in a Sedgwick-Rafter cell, which contains  $1 \times 1$  mm grid divisions, allowing for visualization and quantification under a stereomicroscope, following the protocol described by Martin and Wainwright (2013). The frequency of occurrence (FO) of each dietary item was determined as the proportion of stomachs in which the item was identified relative to the total number of stomachs analyzed (Hyslop 1980). The volume (V) of each item was estimated using the volumetric method described by Hellawell and Abel (1971) and Hyslop (1980). Based on these values, we calculated a modified alimentary index (IAI) for each species, excluding empty stomachs, as proposed by Kawakami and Vazzoler (1980). The obtained proportions were rounded to 0.1% and expressed as percentages. Additionally, we calculated the mean and standard deviation of the proportions of prey items consumed by each species. Dietary items were identified based on partially digested remains, including exoskeletal fragments, plant material, and organic matter. To facilitate analysis, all prey items were classified into taxonomic and functional categories based on size, shape, and movement patterns, including insect larvae, plant material, insects, crustaceans, zooplankton, worms, fish, and detritus (Table 3).

**Table 3.** (a) Stomach content analysis of Characiformes fishes from Mata de Itamacaoca (dry season 2019; N = 8 specimens), showing dietary composition by: frequency of occurrence (F%), volumetric proportion (V%), and Index of Alimentary Importance (IAI). Food items are categorized by taxonomic group, with dominant resources (IAI) indicating key dietary components. (b) Relative contribution of autochthonous and allochthonous food resources to the diet of Characiformes assemblage in Mata de Itamacaoca during the 2019 dry season, based on the Index of Alimentary Importance (IAI)

(a)

Food items/Groups	Frequency of Occurrence (%)	Volume (%)	IAI
Insects			
Coleoptera	19.653	10.268	4.036
Diptera	9.2455	4.4009	0.8140
Ephemeroptera	4.0462	1.9588	0.1585
Hemiptera	8.6705	5.1450	0.8922
Isoptera	4.0462	1.4471	0.1171
Trichoptera	3.4682	2.0771	0.1440
Insect remains	35.260	13.388	9.4413
Insect larvae			
Coleoptera larvae	7.5144	3.1509	0.4735
Diptera larvae	11.560	6.7482	1.5602
Hemiptera larvae	3.4682	1.6484	0.1143
Trichoptera larvae	0.5780	0.2600	0.0030
Plant material			
Flowers	2.8901	1.4261	0.0824
Seeds	18.497	12.898	4.771
Filamentous algae	7.5144	4.6643	0.7010
Plant remains	26.011	12.079	6.2841
Zooplankton			
Hydracarina	3.4682	0.5327	0.0369
Cladocera	0.5780	0.0209	0.0002
Detritus			
Debris	16.184	7.4193	2.4016

Sediment	10.404	3.9495	0.8218
Fish			
Fish scale	9.2485	2.7751	0.5133
Fish remains	0.5780	0.2516	0.0029
Worms			
Nematodeo	1.7341	1.1694	0.0405
Crustaceans			
Decapoda	4.6242	2.3195	0.2145
(b)			
Origin of food items	Main items included	IAI (%)	
Allochthonous	Adult insects (Coleoptera, Diptera, Ephemeroptera, Hemiptera, Isoptera, Trichoptera, insect remains), flowers, seeds, plant remains	79.5	
Autochthonous	Insect larvae (Coleoptera, Diptera, Hemiptera, Trichoptera), filamentous algae, zooplankton (Hydracarina, Cladocera), detritus (debris, sediment), fish tissues (scales, remains), worms (Nematodea), crustaceans (Decapoda)	20.5	

To assess the trophic organization patterns of Characiformes species, we employed a multivariate approach based on the proportions of dietary items identified in stomach contents. As input data, we used the mean proportions (expressed as percentages) of the following dietary items per species: adult insects, insect larvae, plant material, fish, detritus, crustaceans, worms, and zooplankton.

We performed a non-metric multidimensional scaling (nMDS) ordination using a Bray-Curtis dissimilarity matrix calculated from the proportions of dietary items. The analysis was configured with two dimensions and 3,000 iterations, yielding a final stress value of 0.13, indicating a good representation of the data (Clarke 1993). ANOSIM was used to test the hypothesis that differences in dietary item proportions among species were greater than intraspecific variations. Additionally, we conducted an indicator species analysis using the *indicspecies::multipatt* function in R to determine which dietary components significantly contributed to the stomach contents of each species ( $\alpha = 0.05$ ) (Dufrêne and Legendre 1997; De Cáceres et al. 2010). Indicator values were calculated based on the point-biserial correlation coefficient (r.g) between the proportions of each dietary item and species occurrence.

To investigate dietary similarity patterns among species, we performed a hierarchical clustering analysis using the UPGMA (Unweighted Pair Group Method with Arithmetic Mean) method, based on trophic niche overlap (Pianka 1973). Proportional dietary data were standardized using Z-score transformation (scale function). Trophic similarity between species pairs was quantified using the modified Pianka index (Pianka 1973), calculated as:

$$O_{ij} = \frac{\sum_{k=1}^n (p_{ik} * p_{jk})}{\sqrt{\sum_{k=1}^n p_{ik}^2 * \sum_{k=1}^n p_{jk}^2}}$$

Where  $p_{ik}$  e  $p_{jk}$  represent the proportions of dietary item k for species i and j, respectively. This index ranges from 0 (no overlap) to 1 (complete overlap). To convert this similarity measure into a dissimilarity, measure suitable for clustering analysis, we calculated  $D = 1 - O$ .

In addition to its use in clustering analysis, the Pianka index was also applied independently to quantify niche overlap between species pairs. The calculated values were compiled in a matrix to identify species with the highest and lowest trophic overlap (Pianka 1973).

To complement niche overlap analysis, we estimated niche breadth using the Levins' index (Levins 1968), defined as:

$$B = \frac{1}{\sum_{i=1}^n p_i^2}$$

Where:  $B$ : Niche breadth index;  $p_i$ : Proportion of resource  $i$  use relative to the total resources used;  $n$ : Total number of resource categories.

The index was standardized ( $Ba$ ) to a 0–1 scale for cross-species comparisons:

$$Ba = \frac{B - 1}{n - 1}$$

Where  $Ba = 0$ : Specialist (uses only one resource);  $Ba = 1$ : Perfect generalist (equally uses all  $n$  resources).

To summarize dietary patterns at the assemblage level, we fitted linear models (LMs) in R version 4.0.3 (R Core Team 2021) using pooled proportional dietary data from the eight Characiformes species. Proportional data were transformed using the arcsine square root to improve variance homogeneity and normality (Zar 2010; Warton and Hui 2011). The models were used descriptively to evaluate whether the mean proportional contribution of major food categories differed from zero, rather than to test interspecific differences. Model coefficients were therefore interpreted as summaries of assemblage-level dietary composition.

To identify significant differences in dietary proportions among Characiformes species, we performed multiple comparisons using the non-parametric Dunn test (Dunn 1964), with Benjamini-Hochberg correction to control the false discovery rate (Benjamini and Hochberg 1995). The analysis was applied to the transformed data (arcsine square root of proportions; Zar 2010) and considered all paired combinations between species, with a significance level of  $\alpha = 0.05$ .

## Functional morphology analyses

To assess the morphological diversity related to trophic resource use, we performed standardized linear measurements on 20 morphological characters associated with feeding, locomotion, and habitat use, following the morphological scheme illustrated in Oliveira et al. (2024, Table S1) (see Table S2, Table 2). All morphological analyses were performed exclusively on adult individuals, using the same 173 specimens analyzed in the dietary assessments (Table 2). Standard length (SL) ranges confirmed that all individuals fell within adult size classes (Table 2). For this, we adapted protocols from Balon et al. (1986), Sibbing and Nagelkerke (2000), and Breda et al. (2005). Measurements were obtained using a digital caliper (precision of 0.01 mm) and a stereomicroscope, ensuring data accuracy.

To isolate shape variation independently of body size, we applied the Mosimann standardization method, calculating the geometric mean of all measurements per individual and using this value as a divisor for each character. This approach, preferred in recent comparative analyses, allows for a more robust evaluation of morphological adaptations while maintaining the original proportions between characters (Jungers et al. 1995). The geometric mean (GM) was included as an independent variable in subsequent analyses to represent total body size instead of standard length (SL) (Nawa et al. 2024).

To investigate morphological divergence patterns between species, we conducted a Principal Component Analysis (PCA) on the correlation matrix of the standardized measurements. This multivariate analysis allowed us to identify the axes of greatest morphological variation and assess the overlap in the morphospace between species, revealing patterns of morphological segregation. All analytical procedures were performed in the R environment (version 4.1.0).

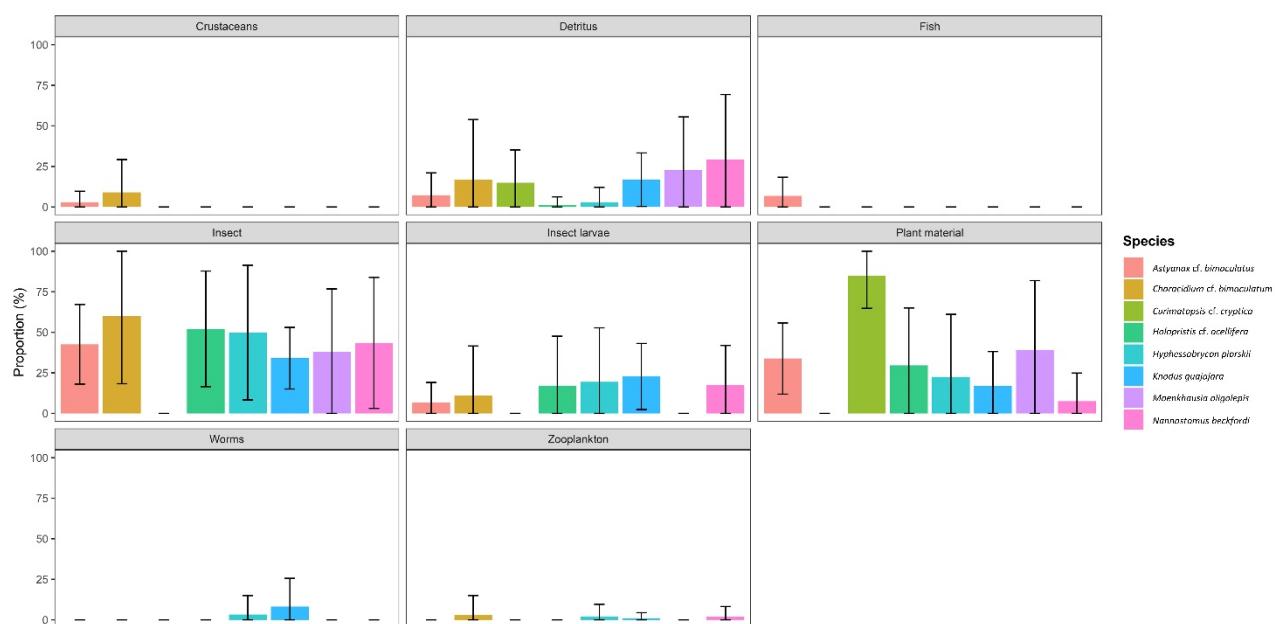
## RESULTS

### Dietary composition

During the dry season, adult insects (61.8%), plant material (54%), and insect larvae (44.1%) dominated the diet of most individuals (Table 3a, b). When dietary items were grouped into autochthonous and allochthonous categories based on their Index of Alimentary Importance (IAI) (Table 3a, b), allochthonous resources (adult insects and terrestrial plant material) accounted for approximately 79.5% of the total dietary importance (Table 3a, b), whereas autochthonous items (insect larvae, algae, zooplankton, detritus, and aquatic invertebrates) contributed the remaining 20.5% (Table 3a, b). Among the species, the highest proportions of adult insect consumption were observed in *Astyanax* cf. *bimaculatus* (42.6%), *Characidium* cf. *bimaculatum* (59.9%), *Holopristis*

*cf. ocellifera* (52.2%), *Nannostomus beckfordi* (43.5%), *Knodus guajajara* (34.1%), and *Hyphessobrycon piorskii* (49.9%) (Fig. 2, Table 3a, b). In contrast, *Curimatopsis cf. cryptica* (85.1%) and *Bario oligolepis* (39.2%) primarily consumed plant material (Fig. 2, Table 3a, b). Some species, such as *Astyanax cf. bimaculatus* and *Characidium cf. bimaculatum*, exhibited higher dietary diversity, incorporating detritus and other resources in smaller proportions (Fig. 2, Table 3a, b).

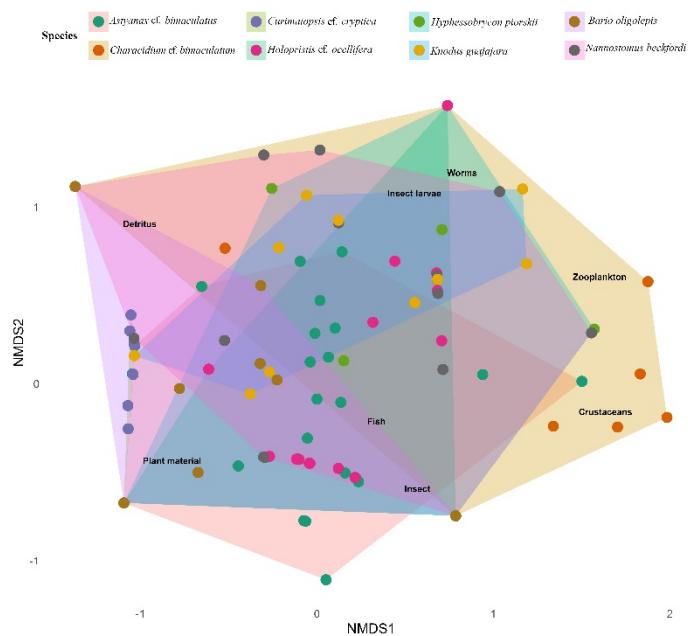
Although some dietary components were rare, such as fish consumption, which was recorded only in *Astyanax cf. bimaculatus* (6.86%), other items like crustaceans were observed in *Astyanax cf. bimaculatus* (2.81%) and *Characidium cf. bimaculatum* (9.02%) (Fig. 2, Table 3a, b). Zooplankton consumption was recorded in *Characidium cf. bimaculatum* (2.97%), *Hyphessobrycon piorskii* (2.15%), *Knodus guajajara* (1.05%), and *Nannostomus beckfordi* (1.95%) (Fig. 2, Table 3a, b). Additionally, worms were recorded exclusively in *Hyphessobrycon piorskii* (3.35%) and *Knodus guajajara* (8.21%) (Fig. 2, Table 3a, b).



**Fig. 2.** Proportion of food items in the diet of the analyzed species. The graphs show the percentage composition (%) of each food category identified in stomach/intestinal contents.

### Clustering, Similarity, and Indicator Species

The NMDS ordination analysis (stress = 0.13, k = 2) revealed a weak clustering of species based on their dietary components, with considerable overlap among them (Fig. 3). However, a statistically significant difference in diet among species was identified (ANOSIM: R = 0.26,  $p = 0.001$ ).



**Fig. 3.** Non-metric Multidimensional Scaling (NMDS) ordination of dietary overlap among of the eight Characiformes species based on stomach content composition (Bray-Curtis dissimilarity). Convex hulls enclose each species' dietary niche space, with closer positions indicating greater similarity in prey composition. Stress value = 0.13, indicating acceptable representation of multidimensional dietary patterns in 2D space.

The results of the indicator species analysis showed significant associations between species and their dietary categories (Table 4). *Astyanax cf. bimaculatus* was associated with fish consumption ( $p < 0.001$ ), while *Knodus guajajara* was associated with worms ( $p = 0.0104$ ) (Table 4). Species combinations showed specific preferences - crustaceans (*Astyanax cf. bimaculatus* + *Characidium cf. bimaculatum*,  $p = 0.011$ ), insects (*Astyanax cf. bimaculatus* + *Bario oligolepis*,  $p = 0.0009$ ), and plant material (*Astyanax cf. bimaculatus* + *Curimatopsis cf. cryptica* + *Bario oligolepis*,  $p = 0.0001$ ) (Table 4). Larger groups favored insect larvae ( $p = 0.007$ ) and detritus ( $p = 0.0094$ ) (Table 4).

**Table 4.** Results of the indicator species analysis (indicspecies) testing for significant dietary preferences among fish species based on stomach content composition. Bold values indicate the most strongly associated prey items for each predator species

Associated Species Group	Prey Category	Indicator Value (stat)	<i>p</i>
<i>Astyanax cf. bimaculatus</i>	Fish	0.556	<b>0.0001***</b>
<i>Knodus guajajara</i>	Worms	0.385	<b>0.0001***</b>
<i>Astyanax cf. bimaculatus</i> + <i>Characidium cf. bimaculatum</i>	Crustaceans	0.364	<b>0.0104*</b>
<i>Astyanax cf. bimaculatus</i> + <i>Curimatopsis cf. cryptica</i> + <i>Bario oligolepis</i>	Plant material	0.532	<b>0.0001***</b>
<i>Astyanax cf. bimaculatus</i> + <i>Bario oligolepis</i>	Insects	0.426	<b>0.0009**</b>
<i>Astyanax cf. bimaculatus</i> + <i>Hypseobrycon piorskii</i> + <i>Knodus guajajara</i> + <i>Nannostomus beckfordi</i>	Insects larvae	0.361	<b>0.007**</b>
<i>Astyanax cf. bimaculatus</i> + <i>Curimatopsis cf. cryptica</i> + <i>Knodus guajajara</i> + <i>Bario oligolepis</i> + <i>Nannostomus beckfordi</i>	Detritus	0.354	<b>0.0094**</b>

## Trophic structure and variation in trophic resource use

The Levins' index ranged from  $Ba = 0.132$  for *Curimatopsis cf. cryptica* to  $Ba = 0.593$  for *Knodus guajajara*, with *Hyphessobrycon piorskii* ( $Ba = 0.577$ ) and *Astyanax cf. bimaculatus* ( $Ba = 0.562$ ) exhibiting the highest values (Table 5). The species utilized between two (*Curimatopsis cf. cryptica*) and six food resources, with *Characidium cf. bimaculatum* and *Nannostomus beckfordi* displaying intermediate values ( $Ba \approx 0.478$ ) (Table 5). *Bario oligolepis* ( $Ba = 0.268$ ) and *Holopristis cf. ocellifera* ( $Ba = 0.372$ ) completed the observed range of variation (Table 5).

**Table 5.** Levin's niche breadth measures: prey proportions (rows 1-8), resource count (N), raw ( $B$ ) and standardized ( $Ba$ ) indices

Dietary component	<i>Astyanax cf. bimaculatus</i>	<i>Characidium cf. bimaculatum</i>	<i>Curimatopsis cf. cryptica</i>	<i>Holopristis cf. ocellifera</i>	<i>Hyphessobrycon piorskii</i>	<i>Knodus guajajara</i>	<i>Bario oligolepis</i>	<i>Nannostomus beckfordi</i>
Insects larvae	0.156	0.2061	0	0.1877	0.272	0.1841	0	0.232
Plant material	0.197	0	0.5971	0.2815	0.1786	0.1439	0.3242	0.1787
Insects	0.3149	0.2482	0	0.371	0.2206	0.2274	0.4226	0.2862
Fish	0.0727	0	0	0	0	0	0	0
Detritus	0.1025	0.1248	0.4029	0.1598	0.1294	0.1319	0.2532	0.242
Crustaceans	0.1568	0.315	0	0	0	0	0	0
Worms	0	0	0	0	0.1605	0.26	0	0
Zooplankton	0	0.1058	0	0	0.0388	0.0528	0	0.0611
N	6	5	2	4	6	6	3	5
$B$	4.933	4.346	1.927	3.601	5.036	5.153	2.875	4.349
$Ba$	0.562	0.478	0.132	0.372	0.577	0.593	0.268	0.478

Dietary niche overlap varied substantially among species (Pianka index: 0.20–1.68) (Table 6). The lowest overlap occurred between *Hyphessobrycon piorskii* and *Knodus guajajara* (0.20), followed by *Curimatopsis cf. cryptica* and *Holopristis cf. ocellifera* (0.72) (Table 6). Conversely, several species pairs showed high overlap (>1.4), particularly *Bario oligolepis* with *Characidium cf. bimaculatum* (1.68) and *Astyanax cf. bimaculatus* with *Nannostomus beckfordi* (1.67) (Table 6). *Curimatopsis cf. cryptica* exhibited low to moderate overlap with most species (0.71–1.42) (Table 6).

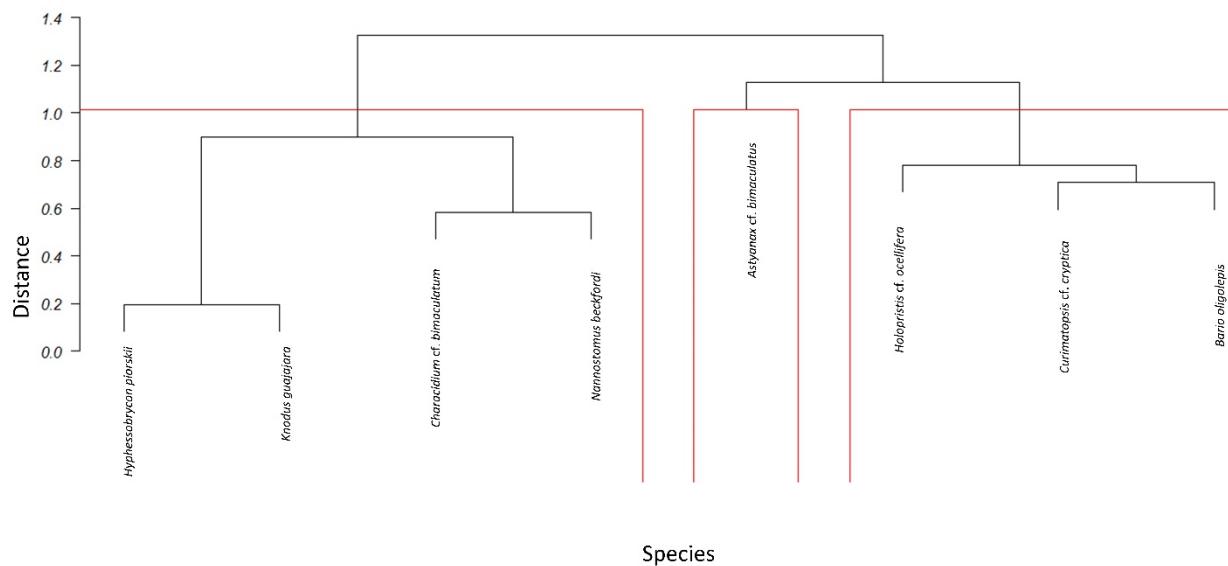
**Table 6.** Pianka's measure of niche overlap (Pianka 1973) among Characiformes species from Mata de Itamacaoca. Values range from 0-1, with 0 being no niche overlap and 1 being complete niche overlap

Species	<i>Astyanax cf. bimaculatus</i>	<i>Characidium cf. bimaculatum</i>	<i>Curimatopsis cf. cryptica</i>	<i>Holopristis cf. ocellifera</i>	<i>Hyphessobrycon piorskii</i>	<i>Knodus guajajara</i>	<i>Bario oligolepis</i>	<i>Nannostomus beckfordi</i>
<i>Characidium cf. bimaculatum</i>	1.1311	1.0000						
<i>Curimatopsis cf. cryptica</i>	1.4241	1.3096	1.0000					
<i>Holopristis cf. ocellifera</i>	1.0375	1.4281	0.7242	1.0000				
<i>Hyphessobrycon piorskii</i>	1.2865	0.9031	1.4173	1.0622	1.0000			
<i>Knodus guajajara</i>	1.4006	0.8876	1.3554	1.4127	0.1959	1.0000		
<i>Bario oligolepis</i>	0.9190	1.6867	0.7074	0.8392	1.6209	1.411	1.0000	
<i>Nannostomus beckfordi</i>	1.6715	0.5843	0.7844	0.9553	0.8329	0.9763	1.3218	1.0000

Pairwise comparisons of species' diets revealed significant differences ( $p < 0.05$ , Benjamini-Hochberg adjusted) in feeding composition among most analyzed pairs. *Astyanax cf. bimaculatus* showed significantly distinct dietary patterns compared to all other species except *Knodus guajajara* ( $p = 0.483$ ) (Table 7). Conversely, *K. guajajara* exhibited pronounced dietary differentiation from most sympatric species, including *Characidium cf. bimaculatum* ( $p < 0.001$ ), *Curimatopsis cf. cryptica* ( $p < 0.001$ ), *Holopristis cf. ocellifera* ( $p = 0.0003$ ), *Hyphessobrycon piorskii* ( $p = 0.001$ ), *Bario oligolepis* ( $p = 0.020$ ), and *Nannostomus beckfordi* ( $p = 0.003$ ) (Table 7). The cluster analysis based on the eight prey categories formed three distinct groups: (1) *Hyphessobrycon piorskii*, *Knodus guajajara*, *Characidium cf. bimaculatum*, and *Nannostomus beckfordi*; (2) *Astyanax cf. bimaculatus*; and (3) *Holopristis cf. ocellifera*, *Curimatopsis cf. cryptica* and *Bario oligolepis* (Fig. 4).

**Table 7.** Mean comparisons between groups adjusted using the Benjamini-Hochberg method. The table displays pairwise mean differences and adjusted  $p$ -values among species groups. ns (not significant). Significant results ( $p \leq 0.05$ ) indicate substantial differences between species pairs

Group 1	Group 2	Mean Difference	Adjusted $p$ -value	Significance
<i>Astyanax cf. bimaculatus</i>	<i>Characidium cf. bimaculatum</i>	-6.097	0.001	**
	<i>Curimatopsis cf. cryptica</i>	-5.337	0.001	**
	<i>Holopristis cf. ocellifera</i>	-4.829	0.001	**
	<i>Hyphessobrycon piorskii</i>	-4.007	0.0001	***
	<i>Knodus guajajara</i>	-0.043	0.483	ns
	<i>Bario oligolepis</i>	-2.776	0.007	**
	<i>Nannostomus beckfordi</i>	-3.866	0.0002	***
<i>Characidium cf. bimaculatum</i>	<i>Curimatopsis cf. cryptica</i>	0.238	0.437	ns
	<i>Holopristis cf. ocellifera</i>	1.689	0.080	ns
	<i>Hyphessobrycon piorskii</i>	0.979	0.241	ns
	<i>Knodus guajajara</i>	5.097	0.001	**
	<i>Bario oligolepis</i>	1.969	0.049	*
	<i>Nannostomus beckfordi</i>	2.154	0.034	*
<i>Curimatopsis cf. cryptica</i>	<i>Holopristis cf. ocellifera</i>	1.319	0.146	ns
	<i>Hyphessobrycon piorskii</i>	0.723	0.299	ns
	<i>Knodus guajajara</i>	4.562	0.001	**
	<i>Bario oligolepis</i>	1.674	0.078	ns
	<i>Nannostomus beckfordi</i>	1.774	0.071	ns
<i>Holopristis cf. ocellifera</i>	<i>Hyphessobrycon piorskii</i>	-0.411	0.381	ns
	<i>Knodus guajajara</i>	3.862	0.0002	***
	<i>Bario oligolepis</i>	0.669	0.307	ns
	<i>Nannostomus beckfordi</i>	0.590	0.324	ns
<i>Hyphessobrycon piorskii</i>	<i>Knodus guajajara</i>	3.495	0.0007	***
	<i>Bario oligolepis</i>	0.912	0.253	ns
	<i>Nannostomus beckfordi</i>	0.866	0.258	ns
<i>Knodus guajajara</i>	<i>Bario oligolepis</i>	-2.424	0.018	*
	<i>Nannostomus beckfordi</i>	-3.154	0.002	**
<i>Bario oligolepis</i>	<i>Nannostomus beckfordi</i>	-0.189	0.441	ns



**Fig. 4.** Dendrogram from cluster analysis on Index of Trophic similarity between species pairs quantified using the modified Pianka index (Pianka 1973) for the eight examined Characiformes fish species in Mata de Itamacaoca, dry season.

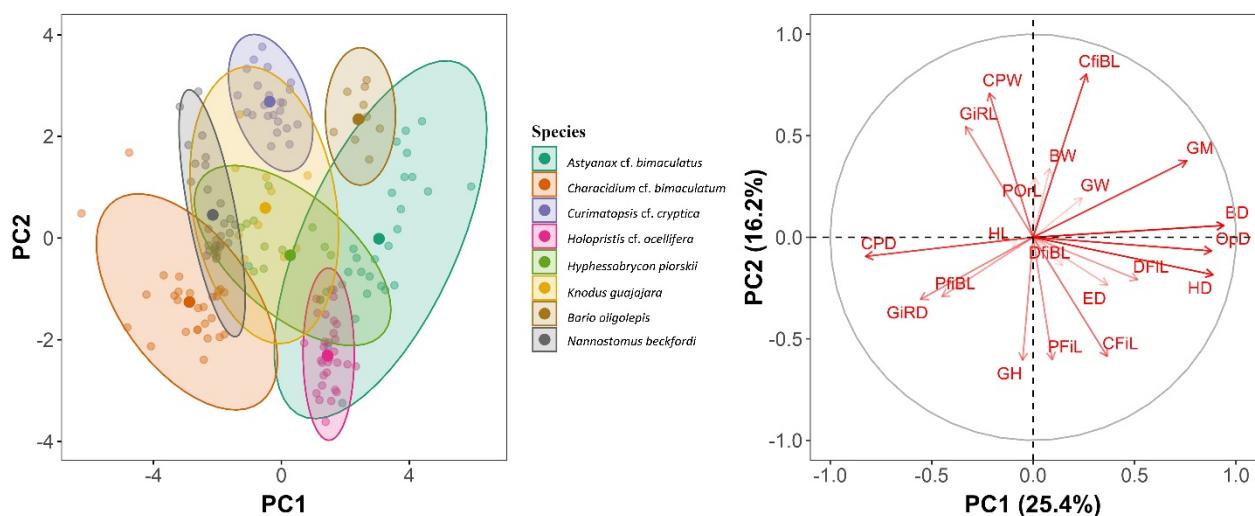
The linear model indicated that the overall mean dietary proportion differed from zero ( $\beta = 0.605$ ;  $p = 0.001$ ; Table 8). Among food categories, insects showed a significant positive coefficient ( $\beta = 0.368$ ;  $p = 0.011$ ; Table 8). Whereas plant material exhibited a marginally significant contribution ( $\beta = 0.285$ ;  $p = 0.051$ ; Table 8). Other food categories, including detritus, fish, insect larvae, worms, and zooplankton, did not differ significantly from zero ( $p > 0.05$ ; Table 8).

**Table 8.** Table 8. Results of linear models (LMs) summarizing assemblage-level dietary composition of Characiformes during the dry season. The table presents estimated coefficients, standard errors, t-values, and significance levels for major food categories. Model coefficients indicate whether the mean proportional contribution of each food category differs from zero. Proportional data were variance-stabilized using an arcsine square root transformation. ms = marginally significant. Statistically significant predictors ( $p < 0.05$ ) are shown in bold

Coefficients	Estimate	Std. Error	T value	<i>p</i>
Intercept	0.605	0.138	4.358	<b>0.001***</b>
Detritus	0.155	0.151	1.028	0.305
Fish	-0.194	0.186	-1.043	0.298
Insect	0.368	0.144	2.545	<b>0.011*</b>
Insect larvae	0.147	0.152	0.963	0.335
Plant material	0.285	0.146	1.955	<b>0.051 ms</b>
Worms	0.087	0.266	0.328	0.743
Zooplankton	-0.116	0.212	-0.550	0.582

## Morphological Variation

The Principal Component Analysis (PCA) explained 41.6% of the total variance, with the first two principal components (PC1 = 25.4%; PC2 = 16.2%) accounting for most of this variance (Fig. 5). Species distribution in the morphological space revealed distinct groupings. *Astyanax* cf. *bimaculatus* was primarily influenced by Caudal fin length (CFiL), while *Characidium* cf. *bimaculatum* was determined by Caudal peduncle depth (CPD) (Fig. 5). For *Curimatopsis* cf. *cryptica*, the most important variable was Body depth (BD), whereas *Holopristis* s cf. *ocellifera* was more influenced by Body width (BW) (Fig. 5). *Hyphessobrycon piorskii* had Head depth (HD) as the predominant variable, while *Knodus guajajara* was influenced by Eye diameter (ED) (Fig. 5). In *Bario oligolepis*, Dorsal fin length (DFiL) had the greatest impact, while *Nannostomus beckfordi* was influenced by Pectoral fin length (PFI). *Bario oligolepis* was influenced by Oral gape width (GW) (Fig. 5).



**Fig. 5.** Biplot of Principal Component Analysis (PCA) of morphological trait space between Characiformes species; and variable loadings on the PC axes.

## DISCUSSION

Here, we present the results of the trophic ecology and morphological analyses of Characiformes species inhabiting the Mata de Itamacaoca, a protected area within the middle Munim River Basin, Maranhão, Brazil. The study was conducted during the dry season and focused on the stomach contents and morphological traits of eight fish species from four different families:

Acestrorhampidae (*Astyanax* cf. *bimaculatus*, *Bario oligolepis*, *Holopristis* cf. *ocellifera*, and *Hyphessobrycon piorskii*), Stevardiidae (*Knodus guajajara*), Lebiasinidae (*Nannostomus beckfordi*), Crenuchidae (*Characidium* cf. *bimaculatum*), and Curimatidae (*Curimatopsis* cf. *cryptica*). Despite the protected status of the area, the presence of urban influences, such as illegal resource extraction, agricultural burning practices, urban encroachment, and inadequate enforcement of conservation measures, highlights the importance of understanding the ecological dynamics of these fish communities (Oliveira et al. 2020 2024). The analyses revealed significant dietary and morphological adaptations, revealing into the mechanisms that allow these species to coexist in a spatially limited and environmentally sensitive habitat during the dry season. Although seasonal hydrological fluctuations broadly influence neotropical aquatic ecosystems, our findings highlight the specific ecological dynamics occurring during the dry season, a critical period of resource scarcity and intensified biotic interactions (Pelage et al. 2022; Londe et al. 2024). While some species presented relatively low sample sizes (e.g., *Knodus guajajara*, *Bario oligolepis*), these numbers are consistent with their observed rarity in the field during the dry season. We interpret these values as biologically meaningful, as they reflect true patterns of local abundance rather than sampling bias.

At the assemblage level, dietary patterns during the dry season were characterized by the predominance of insects and, marginally, plant material, as indicated by the linear model analysis (Table 8). This descriptive overview provides a community-scale context for the morphological patterns discussed below. Although the first two PCA axes accounted for a moderate proportion of total variance (41.6%), such values are common in multivariate ecomorphological datasets that include numerous correlated morphometric traits (Gatz 1979; Winemiller 1991; Jolliffe 2011; Zelditch et al. 2012; Oliveira et al. 2024). Despite this, the PCA revealed clear species-level segregation in morphospace, indicating consistent morphological divergence related to trophic structure. Morphological adaptations among species reflects their feeding preferences: *Astyanax* cf. *bimaculatus*, with a long caudal fin, captures mobile prey (Balon et al. 1986; Breda et al. 2005); *Characidium* cf. *bimaculatum*, with a deep caudal peduncle, enhances burst impulse for insectivory (Sibbing and Nagelkerke 2000); *Curimatopsis* cf. *cryptica*, with a deep body, improves maneuverability (Balon et al. 1986); *Holopristis* cf. *ocellifera*, with a wide body, adapts to vertical movements (Balon et al. 1986); *Hyphessobrycon piorskii*, with a high head, has a varied diet; *Knodus guajajara*, with large eyes, aids in benthic prey detection (Balon et al. 1986); *Bario oligolepis*, with a long dorsal fin, processes vegetation efficiently (Balon et al. 1986; Breda et al. 2005); and *Nannostomus beckfordi*, with extended pectoral fins, controls propulsion (Balon et al. 1986; Breda et al. 2005). Insectivory in *Astyanax* cf. *bimaculatus*, *Characidium* cf. *bimaculatum*, and *Hyphessobrycon piorskii* aligns with Neotropical floodplain patterns (Petry et al. 2011; Esteves

et al. 2021), while phytophagy in *Curimatopsis* cf. *cryptica* (85.07%) and *Bario oligolepis* (39.24%) reflects trophic plasticity (Goulding 1980; Vanni et al. 2006; Medeiros et al. 2014; Allan et al. 2021). Trophic segregation between euryphagous (e.g., *Astyanax* cf. *bimaculatus*) and stenophagous species (e.g., *Knodus guajajara*) supports the "limiting similarity" paradigm (Abrams 1983; Duarte et al. 2022), promoting niche partitioning and reducing competition in seasonal ecosystems (Abrams 1983; Pelage et al. 2022; Londe et al. 2024; Pastore et al. 2021; Zhang et al. 2024).

The consistency between our results and those of Oliveira et al. (2024), conducted in the same area but without accounting for dry season, underscores the significance of insects and plant material as key resources for Characiformes species in the Mata de Itamacaoca during the dry season. *Astyanax* cf. *bimaculatus* diet was characterized by fish and crustaceans in our study, whereas data from Oliveira et al. (2024) emphasized seed intake thus reflecting dry season abundance of resources. Similarly, *Hyphessobrycon piorskii* displayed the presence of worms in our analysis, a dietary component not previously recorded. These discrepancies may reflect dry season fluctuations in resource availability or dietary plasticity, a phenomenon frequently observed in fish inhabiting seasonally dynamic environments, particularly during the dry season (Keller et al. 2019). Nevertheless, the consistent consumption of insects by *Characidium* cf. *bimaculatum* and plant material by *Holopristis* cf. *ocellifera* suggests that these resources play a fundamental role in the trophic ecology of Characiformes species in the Mata de Itamacaoca regardless of environmental variability.

Although species-specific trophic ecology studies were not available for most of the taxa analyzed, we compared our findings with the general trophic patterns reported for their respective genera. Our results generally align with these broader patterns, although notable species-specific differences emerged. For instance, while literature suggests that species of the genera *Knodus* Eigenmann 1911 and *Hyphessobrycon* Durbin 1908 are typically generalist insectivores (Ceneviva-Bastos and Casatti 2007; Prado et al. 2016; Benone et al. 2020), we recorded high insectivory in *Knodus guajajara* (34.1% adult insects) and *Hyphessobrycon piorskii* (49.9%), but also observed niche diversification, such as *Hyphessobrycon piorskii* consumption of worms (3.4%), a resource rarely mentioned in prior studies. Similarly, *Holopristis* cf. *ocellifera* (52.2% insects) and *Bario oligolepis* (39.2% plant material) matched the insectivorous tendency described for their genera (Castro 1999; Graciolli et al. 2003), although *Bario oligolepis* reliance on plant matter was unexpectedly high. *Astyanax* cf. *bimaculatus* and *Characidium* cf. *bimaculatum* exhibited the generalist omnivory documented in earlier work (Casatti et al., 2001; Silva-Camacho et al., 2014), including detritus and crustaceans, but in our data, *A. cf. bimaculatus* also consumed fish remains (6.9%), a trophic behavior less frequently reported for the genus. Both species showed elevated insectivory (42.6% and 59.9%, respectively), surpassing values commonly described in the

literature. *Nannostomus beckfordi*, consistent with the varied diet described for its genus (Silva 1993), also showed high insectivory (43.5%), while incorporating zooplankton and detritus. The most striking divergence was observed in *Curimatopsis* cf. *cryptica*, which predominantly consumed plant material (85.1%) rather than the fine organic matter commonly reported for the genus (Brejão et al. 2013).

These findings corroborate the well-established paradigm that morphological traits are critical determinants of trophic niche specialization, facilitating the efficient exploitation of specific resources through adaptive divergence (Gatz 1979; Sibbing and Nagelkerke 2000; Novakowski et al. 2016). Such morphological relationships are particularly pronounced in freshwater ecosystems, where selective pressures drive functional trait diversification, thereby promoting dietary specialization and mitigating niche overlap via resource partitioning (Ferry-Graham et al. 2002; Montaña and Winemiller 2013; Montaña et al. 2020; Paz Cardozo et al. 2021). The observed congruence between morphology and diet aligns with niche theory (Hutchinson 1957; Chase and Leibold 2009), which posits that phenotypic divergence reduces interspecific competition by enabling differential resource acquisition (Breda et al. 2005; Oliveira et al. 2024). However, the presence of dietary overlap among morphologically distinct species suggests that niche differentiation may also be mediated by non-morphological mechanisms (Chesson 2000; Leibold and McPeek 2006). These could include behavioral plasticity (Gomiero et al. 2010; Garcia et al. 2020), temporal or microhabitat segregation (Schoener 1974; Brandão-Gonçalves and Sebastien 2013), or differential prey selectivity driven by foraging strategies (Lubich et al. 2024). Such compensatory mechanisms may stabilize coexistence in high-diversity assemblages, underscoring the multidimensional nature of niche partitioning (Chesson 2000; Leibold and McPeek 2006). Future studies should integrate functional morphology with spatiotemporal foraging data to disentangle the relative contributions of these factors in structuring trophic interactions.

## CONCLUSIONS

Finally, the ecological implications of resource partitioning and interspecific competition are evident in the coexistence strategies adopted by these species. The observed dietary plasticity, combined with morphological adaptations, suggests that dry season changes in resource availability drive adaptive feeding behaviors that minimize direct competition. This finding supports the hypothesis that environmental dry season acts as a selective pressure, shaping trophic interactions and promoting species coexistence (Bloomfield et al. 2022). However, the proximity of the Mata de Itamacaoca to urban areas raises concerns about anthropogenic disturbances, such as habitat

degradation and water quality deterioration, which could disrupt the delicate balance of resource availability and trophic dynamics (Daufresne and Boet 2007; Matono et al. 2014; Iacarella et al. 2018; Candolin and Rahman 2023). In this context, our study has important conservation implications by identifying functionally vulnerable guilds (*e.g.*, species with restricted diets), establishing baseline data for long-term monitoring, and highlighting critical microhabitats for conservation. Effective protection of this ecosystem thus requires strategies that consider both natural dry season ecological processes and cumulative anthropogenic impacts, integrating aquatic connectivity and the maintenance of habitat heterogeneity.

**Acknowledgments:** We thank CAPES - Fundação Coordenação de Aperfeiçoamento de Pessoal de Nível Superior, Brazil (Finance Code 001), and CNPq - Conselho Nacional de Desenvolvimento Científico e Tecnológico, Brazil for the fellowships: CNPq grants no. 307974/2021-9 and no. 306490/2024-2 to FPO.; CAPES grant no. 88887.699722/2022-00 to E.S.O.; and CAPES grant no. 88887.950525/2024-00 to L.O.V. J.S. acknowledges funding from the UKRI Future Leaders Fellowship (Grant Number: MR/X035662/1).

**Author's Contributions:** Substantial contribution in the concept and design of the study: E.S.O.; J.S.; F.P.O. Specimens' identification: L.O.V.; F.P.O. Contribution to data collection: E.S.O.; L.O.V.; F.P.O. Contribution to data analysis and interpretation: J.S.; E.S.O.; F.P.O. Language revision: J.S. Map preparation: L.O.V. Project financing: F.P.O. Contribution to manuscript preparation: E.S.O.; L.O. V.; J.S.; F.P.O. Contribution to critical revision, adding intellectual content: E.S.O.; L.O.V.; J.S.; F.P.O.

**Competing interests:** The authors declare no conflict of interest. The funders had no role in the design of the study; in the collection, analyses, or interpretation of data; in the writing of the manuscript; or in the decision to publish the results.

**Availability of data and materials:** The data presented in this study are available on request from the corresponding author.

**Consent for publication:** Not applicable.

**Ethics approval and consent to participate:** Not applicable.

## REFERENCES

Abrams P. 1983. The theory of limiting similarity. *Annu Rev Ecol Syst* **14**:359–376.

Abreu JMS, Craig JM, Albert JS, Piorski NM. 2019. Historical biogeography of fishes from coastal basins of Maranhão State, northeastern Brazil. *Neotrop Ichthyol* **17**:1–10. doi:10.1590/1982-0224-20180156.

Albert JS, Tagliacollo VA, Dagosta F. 2020. Diversification of Neotropical freshwater fishes. *Annu Rev Ecol Syst* **51**:27–53. doi:10.1146/annurev-ecolsys-011620-031032.

Alho CJ, Silva JS. 2012. Effects of severe floods and droughts on wildlife of the Pantanal wetland (Brazil)—a review. *Animals* **2**:591–610. doi:10.3390/ani2040591.

Allan JD, Castillo MM, Capps KA. 2021. Energy flow and nutrient cycling in aquatic communities. In: Castillo MM, Capps KA (eds), *Stream ecology: structure and function of running waters*. JD. Springer International Publishing, pp. 357–381. doi:10.1007/978-3-030-61286-3\_12.

Arthington AH, Balcombe SR. 2011. Extreme flow variability and the ‘boom and bust’ ecology of fish in arid-zone floodplain rivers: a case history with implications for environmental flows, conservation and management. *Ecohydrology* **4**:708–720. doi:10.1002/eco.221.

Balon EK, Crawford SS, Lelek A. 1986. Fish communities of the upper Danube River (Germany, Austria) prior to the new Rhein-Main-Donau connection. *Environmental Biology of Fishes*, **15**:243–271. Available at: <https://api.semanticscholar.org/CorpusID:88826468>. Accessed 15 Jan. 2025.

Barbosa JM, Soares EC, Cintra IHA, Hermann M, Araújo AR. 2017. Perfil da ictiofauna da bacia do rio São Francisco/Profile of the fish fauna of the São Francisco River basin. *Acta of Fisheries and Aquatic Resources* **5**:70–90. doi:10.2312/ActaFish.2017.5.1.70-90.

Barreto AP, Aranha JM. 2006. Diet of four species of Characiforms in an Atlantic forest stream, Guaraquecaba, Paraná, Brazil. *Revista Brasileira de Zoologia* **23**:779–788. doi:10.1590/S0101-81752006000300023.

Benjamini Y, Hochberg Y. 1995. Controlling the false discovery rate:a practical and powerful approach to multiple testing. *J R Stat Soc Series B Stat Methodol* **57**:289–300. doi:10.1111/j.2517-6161.1995.tb02031.x.

Benone NL, Lobato CM, Soares BE, Montag LFA. 2020. Spatial and temporal variation of the diet of the flag tetra *Hyphessobrycon heterorhabdus* (Characiforms: Characidae) in streams of the Eastern Amazon. *Neotrop Ichthyol* **18**:1–16. doi:10.1590/1982-0224-2020-0078.

Bloomfield EJ, Guzzo MM, Middel TA, Ridgway MS, McMeans BC. 2022. Seasonality can affect ecological interactions between fishes of different thermal guilds. *Front Ecol Evol* **10**:986459. doi:10.3389/fevo.2022.986459.

Bouton N, Seehausen O, Van Alphen JJM. 1997. Resource partitioning among rock-dwelling haplochromines (Pisces:Cichlidae) from Lake Victoria. *Ecol Freshw Fish* **6**:225–240. doi:10.1111/j.1600-0633.1997.tb00165.x.

Bower LM, Winemiller KO. 2019. Fish assemblage convergence along stream environmental gradients: an intercontinental analysis. *Ecography* **42**:1691–1702. doi:10.1111/ecog.04690.

Brandão-Gonçalves L, Sebastien NY. 2013. Feeding activity and influence of intraspecific competition on zooplankton communities by jundiá (*Rhamdia quelen* Quoy and Gaimard, 1824) in laboratory. *Braz J Biol* **73**:765–773. doi:10.1590/S1519-69842013000400012.

Breda L, Fontes E, Goulart E. 2005. Ecomorfologia de locomoção de peixes com enfoque para espécies neotropicais. *Acta Sci Biol Sci* **27**:371–381. doi:10.4025/actascibiolsci.v27i4.1271

Brejão GL, Gerhard P, Zuanon J. 2013. Functional trophic composition of the ichthyofauna of forest streams in eastern Brazilian Amazon. *Neotrop Ichthyol* **11**:361–373. doi:10.1590/S1679-62252013005000002

Burns MD, Sidlauskas BL. 2019. Ancient and contingent body shape diversification in a hyperdiverse continental fish radiation. *Evolution* **73**:569–587. doi:10.1111/evo.13658.

Burns MD. 2021. Adaptation to herbivory and detritivory drives the convergent evolution of large abdominal cavities in a diverse freshwater fish radiation (Otophysi: Characiformes). *Evolution* **75**:688–705. doi:10.1111/evo.14178

Burns MD, Knouft JH, Dillman CB. 2024. The role of abiotic and biotic factors in the unequal body shape diversification of a Gondwanan fish radiation (Otophysi: Characiformes). *Evolution* **78**:253–266. doi:10.1093/evolut/qpad203

Candolin U, Rahman T. 2023. Behavioural responses of fishes to anthropogenic disturbances: adaptive value and ecological consequences. *J Fish Biol* **103**:773–783. doi:10.1111/jfb.15322.

Castro RMC. 1999. Evolução da ictiofauna de riachos sul-americanos: padrões gerais e possíveis processos causais. In: Caramaschi EP, Mazzoni R, Bizerril CRSF, Peres-Neto PR (Eds) *Ecologia de peixes de riachos: estado atual e perspectivas*. Rio de Janeiro: PPGE-UFRJ, pp.139–155. doi:10.4257/oeco.1999.0601.04.

Casatti L, Langeani F, Castro RMC. 2001. Peixes de riacho do Parque Estadual Morro do Diabo, Bacia do Alto Rio Paraná, SP. *Biota Neotropica* **1**:BN00201122001. doi:10.1590/S1676-06032001000100005.

Clarke KR. 1993. Non-parametric multivariate analyses of changes in community structure. *Australian Journal of Ecology* **18**:117–143. doi:10.1111/j.1442-9993.1993.tb00438.x.

Ceneviva-Bastos M, Casatti L. 2007. Feeding opportunism of *Knodus moenkhausii* (Teleostei: Characidae): an abundant species in streams of the northwestern state of São Paulo, Brazil. *Iheringia, Série Zoologia* **97**:7–15. doi:10.1590/S0073-47212007000100002.

Chesson P. 2000. Mechanisms of maintenance of species diversity. *Annu Rev Ecol Syst* **31**:343–366. doi:10.1146/annurev.ecolsys.31.1.343.

Chase JM, Leibold MA. 2009. Ecological niches:linking classical and contemporary approaches. University of Chicago Press.

Correa SB, Winemiller KO. 2014. Niche partitioning among frugivorous fishes in response to fluctuating resources in the Amazonian floodplain forest. *Ecology*, **95**:210–224. doi:10.1890/13-0393.1

Daufresne M, Boet P. 2007. Climate change impacts on structure and diversity of fish communities in rivers. *Glob Chang Biol* **13**:2467–2478. doi:10.1111/j.1365–2486.2007.01449.x

De Andrade FS, Possamai B, Freitas CEDC, Da Silva Batista J, Hoeinghaus DJ et al. 2024. Niche partitioning and seasonality may mediate coexistence of piranha species in Amazonian floodplain lakes. *Hydrobiologia* **851**:4325–4340. doi:10.1007/s10750-024-05536-z.

De Cáceres M, Legendre P, Moretti M. 2010. Improving indicator species analysis by combining groups of sites. *Oikos* **119**:1674–1684. doi:10.1111/j.1600-0706.2010.18334.x.

Decreto Nº 05/2018, 23 de Março de 2018. Dispõe sobre a criação de área de Relevante Interesse Ecológico (Arie) Itamacaoca. Prefeitura Municipal de Chapadinha, Maranhão, MA.

Duarte C, Antão LH, Magurran AE, de Deus CP. 2022. Shifts in fish community composition and structure linked to seasonality in a tropical river. *Freshw Biol* **67**:1789–1800. doi:10.1111/fwb.13975.

Dufrêne M, Legendre P. 1997. Species assemblages and indicator species: the need for a flexible asymmetrical approach. *Ecol Monogr* **67**:345–366. doi:10.1890/00129615(1997)067[0345:SAAIST]2.0.CO;2

Dunn OJ. 1964. Multiple comparisons using rank sums. *Technometrics* **6**:241–252. doi:10.1080/00401706.1964.10490181

Espírito-Santo HM, Zuanon J. 2017. Temporary pools provide stability to fish assemblages in Amazon headwater streams. *Ecol Freshw Fish* **26**:475–483. doi:10.1111/eff.12292

Esteves KE, Aranha JMR, Albrecht MP. 2021. Ecologia trófica de peixes de riacho: uma releitura 20 anos depois. *Oecologia Australis* **25**:282–282. doi:10.4257/oeco.2021.2502.04.

Ferry-Graham LA, Bolnick DI, Wainwright PC. 2002. Using functional morphology to examine the ecology and evolution of specialization. *Integr Comp Biol* **42**:265–277. doi:10.1093/icb/42.2.265.

Fialho AP, Oliveira LG, Tejerina-Garro FL, de Mérona B. 2008. Fish-habitat relationship in a tropical river under anthropogenic influences. *Hydrobiologia* **598**:315–324. doi:10.1007/s10750-007-9165-3.

Fitzgerald DB, Perez MHS, Sousa LM, Gonçalves AP, Py-Daniel LR et al. 2018. Diversity and community structure of rapids-dwelling fishes of the Xingu River: Implications for conservation

amid large-scale hydroelectric development. *Biological Conservation* **222**:104–112.

doi:10.1016/j.biocon.2018.04.002

Fricke, R., Eschmeyer, W.N., Fong, J.D., 2025a. SPECIES BY FAMILY/SUBFAMILY. Electronic version accessed 09 May 2025.

<http://researcharchive.calacademy.org/research/ichthyology/catalog/SpeciesByFamily.asp>

Fricke R, Eschmeyer WN, van der Laan R. (Eds) 2025b. ESCHMEYER'S CATALOG OF FISHES: GENERA, SPECIES, REFERENCES. Electronic version. Accessed 09 May 2025.

Garcia TD, Quirino BA, Pessoa LA, Cardoso ALP, Goulart E. 2020. Differences in ecomorphology and trophic niche segregation of two sympatric heptapterids (Teleostei: Siluriformes). *Acta Sci Biol Sci* **42**:1–12. doi:10.4025/actascibiolsci.v42i1.49835.

Gatz Jr AJ. 1979. Community organization in fishes as indicated by morphological features. *Ecology* **60**:711–718. doi:10.2307/1936608.

Gerking SD. 1994. Feeding Ecology of Fish. Academic Press, San Diego, 416 p.

Gomiero LM, Villares Junior GA, Naous F. 2010. Seasonal and ontogenetic variations in the diet of *Cichla kelberi* Kullander and Ferreira, 2006 introduced in an artificial lake in southeastern Brazil. *Brazilian Journal of Biology* **70**:1033–1037. doi:10.1590/S1519-69842010000500017.

Goulding M. 1980. The fishes and the forest:explorations in Amazonian natural history Berkeley, University of California Press. p. 280.

Graciolli G, Azevedo MA, Melo FAG. 2003. Comparative study of the diet of Glandulocaudinae and Tetragonopterinae (Ostariophysi, Characidae) in a small stream in southern Brazil. *Stud Neotrop Fauna Environ* **38**:95–110. doi:10.1076/snfe.38.2.95.15932

Hellawell JM, Abel R. 1971. A rapid volumetric method for the analysis of the food of fishes. *J Fish Biol* **3**:29–37. doi:10.1111/j.1095-8649.1971.tb05903.x

Hutchinson GE. 1957. Concluding remarks. *Cold Spring Harb Symp Quant Biol* **22**:415–427. doi:10.1101/SQB.1957.022.01.039.

Hyslop EJ. 1980. Stomach contents analysis: a review of methods and their application. *J Fish Biol* **17**:411–429. doi:10.1111/j.1095-8649.1980.tb02775.x.

Iacarella JC, Adamczyk E, Bowen D, Chalifour L, Eger A et al. 2018. Anthropogenic disturbance homogenizes seagrass fish communities. *Glob Chang Biol* **24**:1904–1918. doi:10.1111/gcb.14090.

IMESC (Instituto Maranhense de Estudos Socioeconômicos e Cartográficos). 2021. Boletim Climático do Maranhão. Available at: <http://imesc.ma.gov.br/portal/Home> (last access on 03 Nov. 2025).

Jungers WL, Falsetti AB, Wall CE. 1995. Shape, relative size, and size-adjustments in morphometrics. *Am J Biol Anthropol* **38**:137–161. doi:10.1002/ajpa.1330380608.

Junk WJ, Bayley PB, Sparks RE. 1989. The flood pulse concept in river-floodplain systems. *Canadian Special Publication of Fisheries and Aquatic Sciences* **106**:110–127.

Jolliffe I. 2011. Principal component analysis. In: International encyclopedia of statistical science. Springer, Berlin, Heidelberg, pp. 1094–1096.

Kawakami E, Vazzoler G. 1980. Método gráfico e estimativa de índice alimentar aplicado no estudo de alimentação de peixes. Boletim do Instituto Oceanográfico **29**:205–207. doi:10.1590/S0373-55241980000200043.

Keller K, Allsop Q, Brim Box J, Buckle D, Crook DA et al. 2019. Dry season habitat use of fishes in an Australian tropical river. Scientific Reports **9**:5677. doi:10.1038/s41598-019-41287-x.

Koerber S, Guimarães EC, Brito PS, Bragança PHN, Ottoni FP. 2022. Checklist of the freshwater fishes of Maranhão, Brazil (CLOFFBR-MA). Ichthyological Contributions of Peces Criollos **79**:1–94.

Langerhans RB, Reznick DN. 2010. Ecology and evolution of swimming performance in fishes: predicting evolution with biomechanics. Fish Locomotion: An Eco-Ethological Perspective **200**:248.

Londe V, Prince CM, Flory SL. 2024. Coexistence in long-term managed lakes: Limited evidence of negative impacts of invasive macrophytes on fish communities. Ecological indicators **169**:112931. doi:10.1016/j.ecolind.2024.112931

Levins R. 1968. Evolution in changing environments: Some theoretical explorations. Princeton University Press.

Leibold MA, McPeek MA. 2006. Coexistence of the niche and neutral perspectives in community ecology. Ecology **87**:1399–1410. doi:10.1890/0012-9658(2006)87[1399:COTNAN]2.0.CO;2

Lubich C, Aguiar-Santos J, Corrêa F, Freitas C, Siqueira-Souza FK. 2024. Trophic ecology of *Acestrorhynchus falcirostris* Cuvier, 1819 in island lakes on the lower stretch of the Solimões River, Amazon Basin. Braz J Biol **84**:e253852. doi:10.1590/1519-6984.253852.

Lytle DA, Poff NL. 2004. Adaptation to natural flow regimes. Trends Ecol Evol **19**:94–100. doi:10.1016/j.tree.2003.10.002.

Martin CH, Wainwright PC. 2013. On the measurement of ecological novelty: scale-eating pupfish are separated by 168 my from other scale-eating fishes. PLoS ONE **8**:e71164. doi:10.1371/journal.pone.0071164.

Matono P, Bernardo JM, Costa AM, Ilhéu M. 2014. Fish response to anthropogenic pressures in temporary streams: the importance of environmental drivers. River Res Appl **30**:1281–1295. doi:10.1002/rra.2780.

Medeiros TN, Rocha AA, Santos NC, Severi W. 2014. Influência do nível hidrológico sobre a dieta de *Leporinus reinhardtii* (Characiformes, Anostomidae) em um reservatório do semiárido brasileiro. Iheringia. Série Zoologia **104**:290–298. doi:10.1590/1678-476620141043290298.

Meira BR, Oliveira FR, Lansac-Tôha FM, Segovia BT, Progênio M et al. 2022. The importance of protists as a food resource for *Astyanax lacustris* (Osteichthyes, Characiformes) larvae at different stages of development. *Hydrobiologia* **849**:781–794. doi:10.1007/s10750-021-04734-3.

Mise FT, Fugi R, Pagotto JPA, Goulart E. 2013. The coexistence of endemic species of *Astyanax* (Teleostei: Characidae) is propitiated by ecomorphological and trophic variations. *Biota Neotropica* **13**:21–28. doi:10.1590/S1676-06032013000300001.

Montaña CG, Ou C, Keppeler FW, Winemiller KO. 2020. Functional and trophic diversity of fishes in the Mekong-3S river system: comparison of morphological and isotopic patterns. *Environ Biol Fishes* **103**:185–200. doi:10.1007/s10641-020-00947-y.

Montaña CG, Winemiller KO. 2013. Evolutionary convergence in Neotropical Cichlids and Nearctic centrarchids: evidence from morphology, diet, and stable isotope analysis. *Biol J Linn Soc* **109**:146–164. doi:10.1111/bij.12021.

Nawa N, South J, Ellender BR, Pegg J, Madzivanzira TC, Wasserman RJ. 2024. Complex selection processes on invasive crayfish phenotype at the invasion front of the Zambezi floodplains ecoregion. *Freshw Biol* **69**:1322–1337. doi:10.1111/fwb.14308

Neves MP, Da Silva JC, Baumgartner D, Baumgartner G, Delariva RL. 2018. Is resource partitioning the key? The role of intra-interspecific variation in coexistence among five small endemic fish species (Characidae) in subtropical rivers. *J Fish Biol* **93**:238–249. doi:10.1111/jfb.13662.

Novakowski GC, Hahn NS, Fugi R. 2008. Diet seasonality and food overlap of the fish assemblage in a pantanal pond. *Neotrop Ichthyol* **6**:567–576. doi:10.1590/S1679-62252008000400004.

Novakowski GC, Cassemiro FAS, Hahn NS. 2016. Diet and ecomorphological relationships of four cichlid species from the Cuiabá River basin. *Neotrop Ichthyol* **14**:e150151. doi:10.1590/1982-0224-20150151.

Núcleo Geoambiental – NuGeo. 2016. Bacias hidrográficas e climatologia no Maranhão. Universidade Estadual do Maranhão, São Luís – MA, p. 165.

Oliveira ES, South J, Vieira LO, Oliveira RF, Ottoni FP. 2025. Trophic dynamics and morphometric divergence of coexisting neotropical cichlid species (Teleostei: Cichliformes) in relation to a dam. *Brazilian Journal of Biology* **85**:e293810. doi:10.1590/1519-6984.293810.

Oliveira ES, South J, Guimarães EC, Vieira LO, Campos DS, Ottoni FP. 2024. Characterizing functional morphology and trophic niches in a neotropical Characiforms (Actinopterygii: Teleostei) assemblage in middle Munim River basin, Maranhão, Brazil. *Brazilian Journal of Biology* **26**:e279881. doi:10.1590/1519-6984.279881.

Pastore AI, Barabás G, Bimler MD, Mayfield MM, Miller TE. 2021. The evolution of niche overlap and competitive differences. *Nat Ecol Evol* **5**:330–337. doi:10.1038/s41559-020-01357-2.

Passos ML, Zambrzycki GC, Pereira RS. 2016. Balanço hídrico e classificação climática para uma determinada região de Chapadinha-MA. *Revista Brasileira de Agricultura Irrigada* **10**:758–76. doi:10.7127/rbai.v10n400402

Paz Cardozo AL, Quirino BA, Yofukuji KY, Ferreira Aleixo MH, Fugi R. 2021. Habitat complexity and individual variation in diet and morphology of a fish species associated with macrophytes. *Ecol Freshw Fish* **30**:184–196. doi:10.1111/eff.12527.

Pazin VF, Magnusson WE, Zuanon J, Mendonca FP. 2006. Fish assemblages in temporary ponds adjacent to ‘terra-firme’ streams in Central Amazonia. *Freshw Biol* **51**:1025–1037. doi:10.1111/j.1365–2427.2006.01537.x.

Pelage L, Lucena-Frédu F, Eduardo LN, Le Loc'h F, Bertrand A, Lira AS, Frédou T. 2022. Competing with each other: Fish isotopic niche in two resource availability contexts. *Front Mar Sci* **9**:975091. doi:10.3389/fmars.2022.975091.

Petry AC, Thomaz SM, Esteves FA. 2011. Comunidade de peixes. In: Esteves FA (ed) *Fundamentos de limnologia*, 3rd edn. Rio de Janeiro, Interciênciac, pp. 609–624.

Pianka ER, 1973. The structure of lizard communities. *Annu Rev Ecol Syst* **4**:53–74.

Poff NL, Ward JV. 1989. Implications of streamflow variability and predictability for lotic community structure: a regional analysis of streamflow patterns. *Canadian Journal of Fisheries and Aquatic Sciences* **46**:1805–1818. doi:10.1139/f89-235.

Porter CK, Golcher-Benavides J, Benkman CW. 2022. Seasonal patterns of dietary partitioning in vertebrates. *Ecol Lett* **25**:2463–2475. doi:10.1111/ele.14083.

Prado AVR, Goulart E, Pagotto JPA. 2016. Ecomorphology and use of food resources: inter- and intraspecific relationships of fish fauna associated with macrophyte stands. *Neotrop Ichthyol* **14**:e150140. doi:10.1590/1982-0224-20150140.

Prejs A, Prejs K. 1987. Feeding of tropical freshwater fishes: seasonality in resource availability and resource use. *Oecologia* **71**:397–404. doi:10.1007/BF00379478.

R Core Team. 2021. R: A language and environment for statistical computing. Vienna, Austria:R Foundation for Statistical Computing.

Reis RE, Albert JS, Di Dario F, Mincarone MM, Petry P, Rocha LA. 2016. Fish biodiversity and conservation in South America. *J Fish Biol* **89**:12–47. doi:10.1111/jfb.13016.

Ross ST. 1986. Resource partitioning in fish assemblages: a review of field studies. *Copeia* **1986**:352–388.

Saint-Paul U, Zuanon J, Correa MAV, García M, Fabré NN et al. 2000. Fish communities in central Amazonian white-and blackwater floodplains. *Environ Biol Fishes* **57**:235–250. doi:10.1023/A:1007699130333.

Sibbing FA, Nagelkerke LA. 2000. Resource partitioning by Lake Tana barbs predicted from fish morphometrics and prey characteristics. *Rev Fish Biol Fish* **10**:393–437.  
doi:10.1023/A:1012270422092.

Silva ALG, Martins F, Santos R, Nunes JLS. 2008. Conservação da Reserva da Itamacaoca de Chapadinha/MA. In: Selbach JF, Leite JRSA, eds. *Meio Ambiente no Baixo Parnaíba: Olhos no mundo, pés na região*. São Luís, Brazil:EDUFMA, pp. 97–104.

Silva CPD. 1993. Alimentação e distribuição espacial de algumas espécies de peixes do igarapé do Candirú, Amazonas, Brasil. *Acta Amazonica* **23**:271–285. doi:10.1590/1809-43921993233285.

Silva-Camacho DDS, Santos JNDS, Gomes RDS, Araújo FG. 2014. Ecomorphological relationships among four Characiformes fish species in a tropical reservoir in South-eastern Brazil. *Zoologia (Curitiba)* **31**:28–34. doi:10.1590/S1984-46702014000100004.

Schoener TW. 1974. Resource partitioning in ecological communities: research on how similar species divide resources helps reveal the natural regulation of species diversity. *Science* **185**:27–39.  
doi:10.1126/science.185.4145.27.

Souza AM, & Auricchio P. 2002 Peixes. In: Auricchio P, Salomão MG (Eds) *Técnicas de coleta e preparação de vertebrados para fins científicos e didáticos*. Instituto Pau Brasil de História Natural. São Paulo, pp. 17–42.

Tonella LH, Ruaro R, Daga VS, Garcia DAZ, Vitorino OB et al. 2022. Neotropical Freshwater Fishes: A dataset of occurrence and abundance of freshwater fishes in the Neotropics. *Ecology* **1**:e3713.  
doi:10.1002/ecy.3713

Underwood W, Anthony R. 2020. AVMA Guidelines for the Euthanasia of Animals:2020 Edition. Retrieved 30 March 2020, 2020-01. Available from: <https://www.avma.org/sites/default/files/2020-01/2020-Euthanasia-Final-1-17-20.pdf>. (viewed 01 Nov. 2022)

Vanni MJ, Bowling AM, Dickman EM, Hale RS, Higgins KA et al. 2006. Nutrient cycling by fish supports relatively more primary production as lake productivity increases. *Ecology* **87**:1696–1709.  
doi:10.1890/0012-9658(2006)87[1696:ncbfsr]2.0.co;2.

Vieira LO, Campos DS, Oliveira RF, South J, Coelho MSP et al. 2023. Checklist of the fish fauna of the Munim River Basin, Maranhão, north-eastern Brazil. *Biodivers Data J* **11**:e98632.  
doi:10.3897/BDJ.11.e98632.

Warton DI, Hui FK. 2011. The arcsine is asinine:the analysis of proportions in ecology. *Ecology* **92**:3–10. doi:10.1890/10-0340.1

Winemiller KO. 1991. Ecomorphological diversification in lowland freshwater fish assemblages from five biotic regions. *Ecol Monogr* **61**:343–365. doi:10.2307/2937046.

Zar JH. 2010. *Biostatistical analysis* pearson prentice-hall. Upper Saddle River, NJ.

Zelditch M, Swiderski D, Sheets HD. 2012. Geometric morphometrics for biologists: a primer. academic press, 478pp.

Zhang Y, Li J, Li Y, Tarkan AS, Liu C, Britton JR. 2024. Trophic niche diversity and redundancy across trophic positions in a subtropical river fish assemblage. *Hydrobiologia* **851**:2417–2428.  
doi:10.1007/s10750-0023-05467-1.

## Supplementary materials

**Table S1.** Sample Codes, Species Names, Voucher Numbers, and Number of Individuals Analyzed in this work. (download)

**Table S2.** Traits measured, morphological descriptions and ecological interpretation of Characiformes of the dry season in Mata de Itamacaoca, middle Munim River basin, Chapadinha municipality, State of Maranhão, northeastern Brazil. (download)